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# Data Validation Summary Report for the 100-DR-1 Operable Unit 100-D-Ponds Phase II Sampling

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**SAMPLING** 

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#### **ACRONYMS**

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%D Percent difference
AA Atomic absorption
BFB Bromofluorobenzene
BNA Base/neutral and acid

CCB Continuing calibration blank
CV Coefficient of variation

CCV Continuing calibration verification
CLP Contract laboratory program
CRA CRDL standard for AA

CRDL Contract Required Detection Limit

CRI CRDL standard for ICP

CRII CRDL standard for ICP initial
CRIF CRDL standard for ICP final
CROL Contract required quantitation limit

CVAA Cold vapor atomic absorption

DBC Dibutylchlorendate

DFTPP Decafluorotriphenylphosphine

DQO Data quality objectives

EPA U.S. environmental protection agency GC/MS Gas chromatography/mass spectrometry

GC Gas chromatography

GFAA Graphite furnace atomic absorption GPC Gel permeation chromatography

ICB Initial calibration blank

ICP Inductively coupled plasma emission spectrometry

ICP interference check sample ICS Initial calibration verification **ICV** Instrument detection limit IDL LCS Laboratory control sample Laboratory control sample soil LCSS Laboratory control sample water LCSW Minimum detectable activity MDA Method of standard addition MSA

MS/MSD Matrix spike/matrix spike duplicate

PBW Preparation blank water PCB Polychlorinated biphenyl

PEM Performance evaluation mixture

QA Quality assurance QC Quality control

RDL Required detection limit

RF Response factor

RIC Reconstructed ion chromatogram

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#### 1.0 INTRODUCTION

The information provided in this validation summary report includes data from the chemical analyses of samples from the 100-DR-1 Operable Unit 100-D-Ponds Phase II Sampling Investigation. All of the data from this sampling event and their related quality assurance samples were reviewed and validated to verify that the reported sample results were of sufficient quality to support decisions regarding remedial actions performed at this site.

Sample analyses included volatile organic, semi-volatile organic, pesticide/PCB, metals, general chemistry and radiochemistry. Three volatile organic samples were analyzed by Quanterra Environmental Services (QTES). The volatile organic samples were validated using Westinghouse-Hanford protocols specified in *Data Validation Procedures for Chemical Analyses*, WHC-SD-EN-SPP-002, Rev 2. All volatile organic data were qualified based on this guidance. The table below lists the volatile organic Sample Delivery Groups (SDGs) that were validated for this sampling event.

SDG No.	Matrix	No. of Samples Analyzed	Level of Validation	Parameters
W0429	S	2	D	Volatile Organics
W0430	S	1	D	Volatile Organics

Three semi-volatile organic samples were analyzed by Quanterra Environmental Services (QTES). The semi-volatile organic samples were validated using Westinghouse-Hanford protocols specified in *Data Validation Procedures for Chemical Analyses*, WHC-SD-EN-SPP-002, Rev. 2. All semi-volatile organic data were qualified based on this guidance. The table below lists the semi-volatile organic SDGs that were validated for this sampling event.

SDG No.	Matrix	No. of Samples Analyzed	Level of Validation	Parameters
W0429	S	2	D	Semi-Volatiles
W0430	S	11	D	Semi-Volatiles

Three pesticide/PCB samples were analyzed by Quanterra Environmental Services (QTES). The pesticide/PCB samples were validated using Westinghouse-Hanford protocols specified in *Data Validation Procedures for Chemical Analyses*, WHC-SD-EN-SPP-002, Rev. 2. All pesticide/PCB data were qualified based on this guidance. The table below lists the pesticide/PCB SDGs that were validated for this sampling event.

SDG No.	Matrix	No. of Samples Analyzed	Level of Validation	Parameters
W0429	S	2	D	General Chem
W0430	S	1	D	General Chem

Three samples were analyzed for radiochemical parameters by QTES laboratories. Radiochemistry sample analyses included the following parameters:

- Gross alpha and gross beta determination
- Gamma spectroscopy

The radiochemical samples were validated using the Westinghouse-Hanford protocols specified in *Data Validation Procedures for Radiochemical Analyses*, WHC-SD-EN-SPP-001, Rev. 1. All radiochemical samples were qualified based on this guidance. The table below lists the radiochemistry SDGs that were validated for this sampling event.

SDG No.	Matrix	No. of Samples Analyzed	Level of Validation	Parameters
W0429	S	2	D	Radiochemistry
W0430	S	1	D	Radiochemistry

The following report is broken down into sections for volatile organic, semi-volatile organic, pesticide/PCB, metals, general chemical and radiochemical analyses. Each volatile organic section includes:

- A general validation summary which addresses precision, accuracy, representaiveness, completeness, and comparability;
- Holding times;
- GC/MS tuning and calibration;
- Blanks, including method blanks;
- Analytical accuracy including matrix spike samples, matrix spike duplicates, surrogates and internal standards performance;
- Analytical precision including matrix spike/matrix spike duplicates;
- Compound identification;
- Sample result quantitation, verification and reported detection limits; and
- System performance and overall assessment.

Each semi-volatile section includes:

- A general validation summary which addresses precision, accuracy, representativeness, completeness, and comparability;
- Holding times;

Each radiochemistry section includes:

- A general validation summary which addresses precision, accuracy, representaiveness, completeness, and comparability:
- Holding times;
- Calibrations;
- Blanks, including laboratory, method and field blanks;
- Analytical accuracy including chemical recoveries, matrix spike samples and laboratory control samples;
- Analytical precision including laboratory duplicates, field duplicates and field splits;
- Sample result quantitation, verification and reported detection limits; and
- System performance and overall assessment.

In addition, the appendices include the data summary tables as well as the validated laboratory report forms for volatile organic, semi-volatile organic, pesticide/PCB, metals, general chemistry and radiochemistry analyses.

Data validation personnel added qualifiers to the reported data based on specified data quality objectives. Qualifiers which may be applied by data validators in compliance with the procedures herein are as follows:

- U Indicates the compound or analyte was analyzed for and not detected in the sample. The value reported is the same quantitation limit corrected for sample dilution and moisture content by the laboratory
- UJ Indicates the compound or analyte was analyzed for and not detected in the sample. Due to a QC deficiency identified during the data validation, the associated quantitation limit is an estimate.
- J Indicates the compound or analyte was analyzed for and detected. Due to a QC deficiency identified during the data validation, the associated concentration is an estimate, but the data are usable for decision-making purposes.
- BJ Applied to inorganic analyses only. Indicates the analyte concentration was greater than the IDL but less than the CRDL and is considered an estimated value.
- R Indicates the compound or analyte was analyzed for, detected, and due to an identified QC deficiency, the data are unusable.
- UR Indicates the compound or analyte was analyzed for and not detected in the sample. Additionally, the data is unusable due to an identified QC deficiency.
- NJ Indicates presumptive evidence of a compound at an estimated value. The data may not be valid for some specific applications (i.e., usable for decision-making purposes).

## 2.0 VOLATILE ORGANIC DATA VALIDATION SUMMARY

#### 2.1 SUMMARY

Positive blank contamination was noted for one methylene chloride result in SDG No. W0429 and for one methylene chloride result in SDG No. W0430. Positive blank results were detected in two acetone results in SDG No. W0429. All associated sample results were flagged accordingly.

With the exceptions noted above, the project-specific data quality objectives in terms of precision, accuracy, completeness, representaiveness, and comparability have been met.

#### 2.2 HOLDING TIMES

Analytical holding times were assessed to ascertain whether the holding time requirements were met by the laboratory. The holding time requirements are as follows: Soil samples must be analyzed within 14 days of the date of sample collection.

If holding times are exceeded, but not by >2x the limit, all associated sample results are qualified as estimates and flagged "J" for detects and "UJ" for non-detects. If holding times are exceeded by >2x the limit, all associated detectable sample results are qualified as estimates and flagged "J" and all non-detects are rejected and flagged "UR".

Holding times were met for all samples.

#### 2.3 GC/MS TUNING AND CALIBRATION

Instrument calibration is performed to establish that the GC/MS instrument is capable of producing acceptable and reliable analytical data over a range of concentrations. The initial and continuing calibrations are performed according to CLP protocols and all results must meet validation requirements set by Westinghouse-Hanford (WHC 1992,b). An initial multipoint calibration is performed prior to sample analysis to establish the linear range of the GC/MS instrument. Continuing calibration checks are performed to verify that instrument performance is stable and reproducible on a day-to-day basis.

All initial and continuing calibration results were acceptable.

All matrix spike results were acceptable.

#### 2.5.2 Surrogates

The analyses of surrogate compounds provide a measure of performance for individual samples. Matrix-specific surrogate compound recovery control windows have been established by the EPA CLP program. When a surrogate compound recovery is out of the control window, all positively identified target compounds associated with the unacceptable surrogate recoveries are qualified as estimates and flagged "J". Undetected compounds with surrogate recoveries less than the lower control limit are qualified as having an estimated detection limit and flagged "UJ". Compounds with surrogate recoveries < 10% are qualified as estimates "J" for detects, and "UR" for nondetects. Undetected compounds with surrogate recoveries greater than the upper control limit require no qualification.

All surrogate recovery results were acceptable.

#### 2.5.3 Internal Standards Performance

The evaluation of the internal standards criteria provide a means to assess the stability and sensitivity of the GC/MS system on every analysis. Internal standard area counts must be within the limits of -50% to +100% of the most recent standard. The retention time of the internal standard must not vary by more than +/-30 seconds of the most recent calibration. If area counts for a particular internal standard are outside the control limits or relative retention time criteria are >+/-30 seconds, all associated sample results are qualified as estimates (J for detects, UJ for non-detects). If area counts and retention times are both outside control limits, all non-detect sample results associated with that internal standard are qualified as unusable "UR".

All internal standard recovery results were acceptable.

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were recalculated to ensure that the reported results were accurate. Raw data were examined for anomalies, transcription errors, and reduction errors. The reviewer verified that the results and detection limits fell within the linear range of the instrument.

All sample results and reported detection limits were acceptable.

#### 2.9 SYSTEM PERFORMANCE AND OVERALL ASSESSMENT

A review of instrument continuing calibration information and QC data indicates that instrument performance was adequate. Positive blank contamination was noted in two acetone results and in one methylene chloride result in SDG No. W0429. Positive blank contamination was noted in one methylene chloride result in SDG No. W0430. The associated sample results were flagged accordingly. Contamination, however, was not significantly high enough to affect the usability of the data. All other validated results are considered accurate within the standard error associated with the methods.

All data packages submitted for validation were found to be 100% complete.

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#### 3.0 SEMI-VOLATILE ORGANIC DATA VALIDATION SUMMARY

#### 3.1 SUMMARY

Positive blank contamination was noted for all aldol condensate and butylbenzylphthalate results in both SDGs. Due to an internal standard recovery outside control limits, all results in SDG No. W0429 associated with internal standard perylene-d12 were qualified as estimates. All associated sample results were flagged accordingly.

With the exceptions noted above, the project-specific data quality objectives in terms of precision, accuracy, completeness, representativeness, and comparability have been met.

#### 3.2 HOLDING TIMES

Analytical holding times were assessed to ascertain whether the holding time requirements were met by the laboratory. The holding time requirements are as follows: Soil samples must be extracted within 14 days of the date of sample collection and analyzed within 40 days from the date of extraction.

If holding times are exceeded, but not by > 2x the limit, all associated sample results are qualified as estimates and flagged "J" for detects and "UJ" for non-detects. If holding times are exceeded by > 2x the limit, all associated detectable sample results are qualified as estimates and flagged "J" and all non-detects are rejected and flagged "UR".

Holding times were met for all samples.

#### 3.3 GC/MS TUNING AND CALIBRATION

Instrument calibration is performed to establish that the GC/MS instrument is capable of producing acceptable and reliable analytical data over a range of concentrations. The initial and continuing calibrations are performed according to CLP protocols and all results must meet validation requirements set by Westinghouse-Hanford (WHC 1992,b). An initial multipoint calibration is performed prior to sample analysis to establish the linear range of the GC/MS instrument. Continuing calibration checks are performed to verify that instrument performance is stable and reproducible on a day-to-day basis.

All initial and continuing calibration results were acceptable.

estimates and flagged "UJ". Sample results > 5x the spike concentration require no qualification.

All matrix spike results were acceptable.

#### 3.5.2 Surrogates

The analyses of surrogate compounds provide a measure of performance for individual samples. Matrix-specific surrogate compound recovery control windows have been established by the EPA CLP program. When a surrogate compound recovery is out of the control window, all positively identified target compounds associated with the unacceptable surrogate recoveries are qualified as estimates and flagged "J". Undetected compounds with surrogate recoveries less than the lower control limit are qualified as having an estimated detection limit and flagged "UJ". Compounds with surrogate recoveries < 10% are qualified as estimates "J" for detects, and "UR" for nondetects. Undetected compounds with surrogate recoveries greater than the upper control limit require no qualification.

All surrogate recovery results were acceptable.

#### 3.5.3 Internal Standards Performance

The evaluation of the internal standards criteria provide a means to assess the stability and sensitivity of the GC/MS system on every analysis. Internal standard area counts must be within the limits of -50% to +100% of the most recent standard. The retention time of the internal standard must not vary by more than +/-30 seconds of the most recent calibration. If area counts for a particular internal standard are outside the control limits or relative retention time criteria are >+/-30 seconds, all associated sample results are qualified as estimates (J for detects, UJ for non-detects). If area counts and retention times are both outside control limits, all non-detect sample results associated with that internal standard are qualified as unusable "UR".

Due to an internal standard recovery outside control limits, all results associated with internal standard perylene-d12 in SDG No. W0429 have been qualified as estimates and flagged "UJ/J".

All other internal standard recovery results were acceptable.

the project-specific CRQLs were met. Sample results and reported detection limits were recalculated to ensure that the reported results were accurate. Raw data were examined for anomalies transcription errors, and reduction errors. The reviewer verified that the results and detection limits fell within the linear range of the instrument.

All sample results and reported detection limits were acceptable.

#### 3.9 SYSTEM PERFORMANCE AND OVERALL ASSESSMENT

A review of instrument continuing calibration information and QC data indicates that instrument performance was adequate. Positive blank contamination was noted in all aldol condensate and butylbenzylphthalate results in both SDGs. The associated sample results were flagged accordingly. Due to an internal standard recovery outside control limits, all results in SDG No. W0429 associated with internal standard perylene-d12 were qualified as estimates and flagged "UJ/J". Data flagged "J" indicate the associated concentration is an estimate, but the data are usable for decision making purposes. All other validated results are considered accurate within the standard error associated with the methods.

All data packages submitted for validation were found to be 100% complete.

#### 4.0 PESTICIDE/PCB DATA VALIDATION SUMMARY

#### 4.1 SUMMARY

Due to calibration verification results outside QC limits, twelve compounds in SDG No. W0429 and eleven compounds in SDG No. W0430 were qualified as estimates. All associated sample results were flagged accordingly.

With the exceptions noted above, the project-specific data quality objectives in terms of precision, accuracy, completeness, representativeness, and comparability have been met.

#### 4.2 HOLDING TIMES

Analytical holding times were assessed to ascertain whether the holding time requirements were met by the laboratory. The holding time requirements are as follows: Soil samples must be extracted within 14 days of the date of sample collection and analyzed within 40 days from the date of extraction.

If holding times are exceeded, but not by > 2x the limit, all associated sample results are qualified as estimates and flagged "J" for detects and "UJ" for non-detects. If holding times are exceeded by > 2x the limit, all associated detectable sample results are qualified as estimates and flagged "J" and all non-detects are rejected and flagged "UR".

Holding times were met for all samples.

#### 4.3 GC/MS TUNING AND CALIBRATION

#### 4.3.1 Initial Calibration

The laboratory performed an initial multipoint calibration for all target compounds at the concentration required by SW-846 protocols. The linearity of the initial calibration is established when the %RSD or the calibration factors are <20%. If the RSD is >20%, all detected results are qualified as estimates and flagged "J", and all non-detects are flagged "UJ".

All initial calibration results were acceptable.

compounds and % recoveries must be within established laboratory quality control limits. If spike recoveries are outside control limits, detected sample results <5x the spike concentration are qualified as estimates and flagged "J". Undetected sample results with spike recoveries outside control limits are qualified as estimates and flagged "UJ". Sample results >5x the spike concentration require no qualification.

All matrix spike results were acceptable.

#### 4.5.2 Surrogates

The analyses of surrogate compounds provide a measure of performance for individual samples. Matrix-specific surrogate compound recovery control windows have been established by the laboratory. When a surrogate compound recovery is out of the control window, all positively identified target compounds associated with the unacceptable surrogate recoveries are qualified as estimates and flagged "J". Undetected compounds with surrogate recoveries less than the lower control limit are qualified as having an estimated detection limit and flagged "UJ". Compounds with surrogate recoveries < 10% are qualified as estimates "J" for detects, and "UR" for nondetects. Undetected compounds with surrogate recoveries greater than the upper control limit require no qualification.

All surrogate recovery results were acceptable.

#### 4.6 ANALYTICAL PRECISION

#### 4.6.1 Matrix spike/matrix spike duplicates

Matrix spike/matrix spike duplicate results provide matrix-specific information on the precision of the method for specific target compound classes. Precision is expressed by the RPD between the recoveries of duplicate matrix spike analyses performed on a sample. For soil samples analyzed using SW-846 protocol, results must be within RPD limits of +/-35%. If RPD values are out of specification and the sample concentration is <5x the spike concentration, all associated detected sample results are qualified as estimates and flagged "J". If RPD values are out of specification and the sample concentration is >5x the spike concentration, no qualification is required.

All matrix spike/matrix spike duplicate RPD results were acceptable.

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#### 5.0 METALS DATA VALIDATION SUMMARY

#### 5.1 SUMMARY

Negative blank contamination was detected in one sample. Minor matrix spike recovery problems were noted for three analytes in one sample delivery group. All associated sample results were flagged accordingly.

With the exceptions noted above, the project-specific data quality objectives in terms of precision, accuracy, completeness, representaiveness, and comparability have been met.

#### 5.2 HOLDING TIMES

Analytical holding times for ICP metals and GFAA metals analyses were assessed to ascertain whether the holding time requirements were met by the laboratory. The holding time requirements are as follows: Samples must be analyzed within six months for all metals.

Holding times were met for all samples.

#### 5.3 CALIBRATIONS

Performance of specific instrument quality assurance and quality control procedures, including deficiencies noted during the quality assurance review, are outlined below.

Three calibration standards and a blank were analyzed for arsenic, lead, selenium and thallium by GFAA. The correlation coefficient of a least squares linear regression met the requirements for calibration in all cases.

At least one standard and a blank were analyzed by ICP for all other elements.

The above calibrations were each immediately verified with an ICV standard and a calibration blank. The ICV was prepared from a source independent of the calibration standards, at a mid-calibration range concentration. The ICV percent recovery must fall within the control limits of 90% to 110% for metals analyzed by ICP and GFAA. Calibration linearity near the detection limit was verified with a standard prepared at a concentration near the CRDL.

The ICVs met the recommended control limits in all cases.

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within (2x) the absolute blank value are qualified as estimates and flagged "J". The qualification applies only to results generated between the associated calibration blank and the nearest acceptable calibration blank.

All calibration blank results were acceptable.

#### 5.4.2 Preparation Blanks

At least one preparation blank, consisting of deionized distilled water must be prepared and analyzed with every sample delivery group. In the case of positive blank results, samples with results (in ug/L) of (<5x) the preparation blank value have had their associated values qualified as non-detected and flagged "U". Samples with concentrations of (>5x) the highest blank concentration do not require qualification.

If the absolute value of the negative preparation blank exceeds the CRDL, all associated undetected results are rejected and flagged "UR". All associated detects that are (<10x) the absolute value of the preparation blank result are qualified as estimates and flagged "J". If the sample results are (>10x) the absolute value of the preparation blank, no qualification is necessary. If the absolute value of the negative preparation blank is >1DL and  $\le$ CRDL, all associated non-detected sample results are qualified as estimates and flagged "UJ". All associated detects (<10x) the absolute value of the preparation blank are qualified as estimates and flagged "J".

Due to the presence of a negative preparation blank result, sample number BODMT4 in SDG No. W0430 was flagged "BJ" for beryllium.

All other preparation blank results were acceptable.

#### 5.5 ACCURACY

#### 5.5.1 Matrix Spike Samples

Matrix spike analyses are used to assess the analytical accuracy of the reported data and the effect of the matrix on the ability to accurately quantify sample concentrations. Matrix spike recoveries must fall within the range of 75% to 125%. Samples with a spike recovery of <30% and a sample value below the IDL were rejected and flagged "UR". Samples with a spike recovery of 30% to 74% and a sample result <IDL are qualified "UJ". Samples with a spike recovery of >125% or <75% and a sample result >IDL are qualified "J". All samples with a spike recovery > 125% and a sample result <IDL require no qualification.

#### 5.5.4 Method of Standard Addition (MSA) Results

For all samples whose analytical spike results are outside the 85% to 115% control limit and whose absorbances are >50% of the analytical spike absorbance, an MSA is required. In cases where the MSA correlation coefficient was <0.995, the MSA analysis was repeated once. If the correlation coefficient was still <0.995, samples were qualified as estimates and flagged "J". If a sample required MSA analysis but was not analyzed, all associated data must be qualified as estimates and flagged "J".

All MSA results were acceptable.

#### 5.6 ANALYTICAL PRECISION

#### 5.6.1 Laboratory Duplicate Samples

The laboratory duplicate results assess the precision of the method by measuring a second aliquot of the sample that is treated the same way as the original. If the RPD of the original sample and its duplicate is >35% and the positive sample result is (>5x) the CRDL, the associated sample result is qualified as an estimate and flagged "J". Also, if the difference between the duplicate samples is  $>\pm$  CRDL and the positive sample result is (<5x) the CRDL, the associated sample result is qualified as an estimate and flagged "J".

All laboratory duplicate recovery results were acceptable.

#### 5.6.2 ICP Serial Dilution

The ICP serial dilution is used to determine whether significant physical or chemical interferences exist due to sample matrix. If a sample concentration is  $(\ge 50x)$  the IDL for an analyte and the %D is outside the +10% control limits the associated data must be qualified as estimates and flagged "J".

No ICP serial dilution was analyzed with the SDGs in this report. No data was qualified since SW-846 methods do not require dilution analysis unless sample concentrations are greater than the linear range of the instrument.

#### 5.6.3 Field Duplicates

Field duplicate results are compared using the same guidelines for determining the RPD between a sample and its duplicate. Data are not qualified based on field duplicates.

#### 5.8 SYSTEM PERFORMANCE AND OVERALL ASSESSMENT

A review of instrument continuing calibration information and QC data indicates that instrument performance was adequate. Negative blank contamination was detected in one beryllium sample in SDG No. W0430. The associated sample result was flagged accordingly. Contamination, however, was not significantly high enough to affect the usability of the data. Due to matrix

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spike recovery problems, antimony, manganese and thallium results in SDG No. W0430 were qualified as estimates and flagged (BJ, J, or UJ). Data flagged "J" indicates that the associated concentration is an estimate, but the data are usable for decision making purposes. All other validated results are considered accurate within the standard error associated with the methods.

All data packages submitted for validation were found to be 100% complete.

#### 6.0 GENERAL CHEMISTRY DATA VALIDATION SUMMARY

#### 6.1 SUMMARY

The project-specific data quality objectives in terms of precision, accuracy, completeness, representativeness, and comparability have been met.

#### 6.2 HOLDING TIMES

Analytical holding times for fluoride, sulfate, chloride, sulfide, nitrate, nitrite, phosphate, nitrate/nitrite and TOX were assessed to ascertain whether the holding time requirements were met by the laboratory. The holding time requirements are as follows: 28 days for chloride, fluoride, sulfate, TOX and nitrate/nitrite; seven days for sulfide; and two days for nitrate, nitrite and phosphate.

If holding times are exceeded, but not by (>2x) the limit, all associated sample results are qualified as estimates and flagged "J" for detects and "UJ" for non-detects. If holding times are exceeded by (>2x) the limit, all associated detectable sample results are qualified as estimates and flagged "J" and all non-detects are rejected and flagged "UR".

Holding times for all analytes met QC requirements.

#### 6.3 CALIBRATIONS

#### 6.3.1 Initial Calibration

The following calibration procedures must be conducted:

- At least one blank and three standards were used to establish the ion chromatography, ion selective electrode, and spectrophotometer calibrations prior to sample analysis with a correlation >0.995.
- At least two reference buffers or standards at a high and low concentration were used to calibrate the pH and conductivity meters.

If any of these initial calibration requirements are not met, all associated data are qualified "J" for detects and "UJ" for non-detects.

All initial calibration results were acceptable.

performance criteria for solid LCS samples are established by the manufacturer or the laboratory.

ICV results obtained from the raw data were used to calculate LCS results. All LCS results were found to be acceptable.

#### 6.6 PRECISION

#### 6.6.1 Laboratory Duplicates

The laboratory duplicate sample analyses are used to measure laboratory precision and sample homogeneity. Laboratory duplicate RPDs must fall below 20% for waters and 35% for soils. If an RPD for an aqueous sample is >20% and the sample result is (>5x) the CRDL, all associated detects are qualified as estimates and flagged "J". If the range between duplicate aqueous samples is  $>\pm$  CRDL and the sample result is (<5x) the CRDL, all associated detects are qualified as estimates and flagged "J". If an RPD for soil samples is >35% and the sample result is (>5x) the CRDL, all associated detects are flagged "J". If the range between duplicate soil samples is  $>\pm$  2CRDL and the sample result is (<5x) the CRDL, then all detects are flagged "J".

All laboratory duplicate results were acceptable.

#### 6.6.2 Field Duplicates

Field duplicate sample analyses are used to measure both the lab and field sampling procedure precision. Field duplicate results are compared using the same guidelines for determining the precision between a sample and its duplicate. Data are not qualified based on field duplicates.

There were no field duplicates associated with the subject SDGs.

#### 6.6.3 Field Split Samples

A field split sample is a representative sample from a sampling event that is sent to a third party laboratory. Field split sample results are evaluated by comparing the corresponding sample results to the reference laboratory sample results. Data qualification is not required for field splits.

There were no field splits associated with the subject SDGs.



#### 7.0 RADIOCHEMISTRY DATA VALIDATION SUMMARY

#### 7.1 SUMMARY

Radium-228 results for all samples in SDG No. W0429 were qualified as estimates due to the lack of a duplicate analysis. Uranium-238 results for all samples in SDG No. W0429 were qualified as estimates due to a laboratory duplicate RPD of 39%. All associated sample results were flagged accordingly.

With the exceptions noted above, the project-specific data quality objectives in terms of precision, accuracy, completeness, representaiveness, and comparability have been met.

#### 7.2 HOLDING TIMES AND SAMPLE PREPARATION

Holding times are calculated from Chain-of-Custody forms to determine the validity of the results. The maximum holding time for radiochemical analyses is six months. Tritium sample preparation requires distillation. Tritium samples must be analyzed within seven days of distillation.

All holding times and sample preparation measures were acceptable.

#### 7.3 CALIBRATIONS

Instrument calibration is performed to establish that the counters used to determine radionuclide activities are capable of producing acceptable and reliable analytical data. Each counting system must be factory calibrated at installation and after any maintenance or repair. Calibration consists of an instrument efficiency determination for each applicable radionuclide. Continuing calibration checks are performed to verify that instrument performance is stable and reproducible.

All calibration results, including efficiency checks and background counts, were acceptable.

#### 7.4 LABORATORY BLANKS

Laboratory blank samples are analyzed to determine if positive results are due to laboratory reagent, sample container, or detector contamination. If blank analysis results indicate the presence of an analyte above the MDA, the following qualifiers were applied: All positive sample results (<5x) the highest blank concentration were qualified as estimated; sample results below the MDA were

#### 7.6.2 Field Duplicates

Field duplicate results are compared using the same guidelines for determining the RPD between a sample and its duplicate. Data qualification is not required for field duplicate RPDs.

There were no field duplicates associated with the subject SDGs.

#### 7.6.3 Field Split Samples

A field split sample is a representative sample from a sampling event that is sent to a third party laboratory. The field split sample results are evaluated by comparing the corresponding sample results to the reference laboratory sample results. Data qualification is not required for field splits.

There were no field splits associated with the subject SDGs.

# 7.7 SAMPLE RESULTS QUANTITATION, VERIFICATION AND REPORTED DETECTION LIMITS

Twenty percent of sample results and reported detection limits were recalculated to ensure that the reported results were accurate. Raw data were examined for anomalies, transcription errors, and reduction errors. The MDA for each analyte was assessed to ensure that it met the CRDL.

The reviewer verified that the results and detection limits fell within the linear range of the instrument. All data packages submitted for validation were found to be complete. All sample results and reported detection limits were acceptable.

#### 7.8 SYSTEM PERFORMANCE AND OVERALL ASSESSMENT

A review of instrument continuing calibration information and QC data indicates that instrument performance was adequate for these analyses. Due to the lack of a duplicate analysis, all radium-228 results in SDG No. W0429 were qualified as estimates and flagged "J". All uranium-238 results in SDG No. W0429 were qualified as estimates due to a laboratory duplicate RPD of 39%. Data flagged "J" indicate the associated concentration is an estimate, but the data are usable for decision making purposes. All other validated results are considered accurate within the standard error associated with the methods.

All data packages submitted for validation were found to be 100% complete.

#### 8.0 REFERENCES

34.346.1729

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- EPA, 1988b, Laboratory Data Validation Functional Guidelines for Evaluating Organics Analyses, U.S. Environmental Protection Agency, Washington, D.C.
- EPA, 1988c, EPA Contract Laboratory Program Statement of Work for Inorganics Analyses, Multi-Media, Multi-Concentration, U.S. Environmental Protection Agency, Washington, D.C.
- EPA, 1988d, Laboratory Data Validation Functional Guidelines for Evaluating Inorganics Analyses, U.S. Environmental Protection Agency, Washington, D.C.
- EPA, 1990, EPA Contract Laboratory Program Statement of Work for Inorganic Analyses, Multi-media, Multi-Concentration, U.S. Environmental Protection Agency, Washington, D.C.
- EPA, 1991, EPA Contract Laboratory Program Statement of Work for Organics Analyses, Multi-Media, Multi-Concentration, Environmental Protection Agency, Washington, D.C.
- WHC, 1992a, Data Validation Procedures for Chemical Analyses, WHC-SD-EN-SPP-002, Rev. 2, Westinghouse Hanford Company, October 1993.
- WHC, 1992b, Data Validation Procedure for Radiological Analyses, WHC-SD-EN-SPP-001, Rev. 2, Westinghouse Hanford Company, 1993.

#### **APPENDICES**

# APPENDIX A VOLATILE ORGANIC DATA SUMMARY TABLES

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Project: WESTINGHOUSE-HA	ANFORD			i															
Laboratory: QUANTERRA	1 a a a	15.55																	
Case:	SDG: V			<u></u>												T			
Sample Number		BODMT2		BODMT3												<u> </u>		<u> </u>	
Location		Test Fit #	<u> </u>	Test Pit	<u> 2</u>	<u></u>				<u> </u>						<u> </u>			
Remarks																			
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Analysis Date		01/25/95		01/26/95			1.8		IA	D : 14	- X	B 4	- A	Beauta	<b>一</b>	No. of the	TA.	Danile.	TX
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BHI-00405 Rev. 00

#### APPENDIX B

### **VOLATILE ORGANIC VALIDATED LABORATORY REPORT FORMS**

# VOLATILE ORGANICS ANALYSIS DATA SHEET

BHI-00405 Rev. 00 BODMT2

Lab Name: QUANTERRA MO Contract: 550-56

Lab Code: ITMO Case No.: V34402 SDG No.: W0429

Matrix: (soil/water) SOIL Lab Sample ID: 7344-002

Sample wt/vol: 5.00 (g/mL) G Lab File ID: F0621

Level: (low/med) LOW Date Received: 01/18/95

% Moisture: not dec. 44 Date Analyzed: 01/25/95

Column: (pack/cap) CAP Dilution Factor: 1.0

CAS NO. COMPOUND (ug/L or ug/Kg) UG/KG

CAS NO.	COMPOUND	(ug/L or			Q	
	Chloromethane			18	U	1
	Bromomethane		i	18	U	1
75-01-4	Vinyl Chlori $\overline{\mathtt{de}}$			18	U	
75-00-3	Chloroethane		i	18	U	
75-09-2	Methylene $Ch\overline{lc}$	ride		20	B U	1
67-64-1	Acetone			43	JB/2 N	
75-15-0	Carbon Disulfi	de		9	ប់	1
75-35-4	1,1-Dichloroet	hene		9	U	(
75-34-3	1,1-Dichloroet	hane		9	<b>ט</b>	
540-59-0	1,2-Dichloroet	hene (total)	<del></del> 1	9	U	1
	Chloroform	-	<del></del>	9	U	
107-06-2	1,2-Dichloroet	hane		9	U	1
78-93-3	2-Butanone			180	טן	
71-55-6	1,1,1-Trichlor	oethane	<del></del> 1	9	ט	- (
56-23-5	Carbon Tetrach	loride		9	U	
75-27-4	Bromodichlorom	ethane	<del></del> -1	9	ไซ	
78-87-5	1,2-Dichloropr	opane	<del></del>	و	Ū	1
10061-01-5-	cis-1,3-Dichlo	ropropene	<del></del> 1	9	Ū	- (
79-01-6	Trichloroethen	e	<del></del>	9	Ū	
124-48-1	Dibromochlorom	et hane	<del></del> 1	9	Ū	ĺ
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# TA THE VOLATILE ORGANICS ANALYSIS DATA SHEET

EPA SAMPLE NO. BHI-00405

Rev. 00 BODMT3

Lab Name: QUANTERRA MO Contract: 550-56

Lab Code: ITMO Case No.: V34402 SDG No.: W0429

Matrix: (soil/water) SOIL Lab Sample ID: 7344-004

Sample wt/vol: 5.00 (g/mL) G Lab File ID: F0635

Level: (low/med) LOW Date Received: 01/18/95

% Moisture: not dec. 68 Date Analyzed: 01/26/95

Column: (pack/cap) CAP Dilution Factor: 1.0

CONCENTRATION UNITS:

CAS NO. COMPOUND (ug/L or ug/Kg) UG/KG Q

74-87-3				<del>,</del>
74-83-9	74-87-3	Chloromethane	31	U
75-01-4			31	U
75-00-3			31	U
75-09-2			31	U.
67-64-1			74	B
75-15-0			51	NICE
75-35-4			16	
75-34-31,1-Dichloroethane       16       U         540-59-01,2-Dichloroethene       (total)       16       U         67-66-3			16	ט
540-59-01, 2-Dichloroethene       16       U         67-66-3Chloroform       16       U         107-06-21, 2-Dichloroethane       16       U         78-93-32-Butanone       310       U         71-55-61, 1, 1-Trichloroethane       16       U         56-23-5Carbon Tetrachloride       16       U         75-27-4Bromodichloromethane       16       U         78-87-51, 2-Dichloropropane       16       U         10061-01-5cis-1, 3-Dichloropropene       16       U         79-01-6Trichloroethane       16       U         79-00-51, 1, 2-Trichloroethane       16       U         71-43-2Benzene       16       U         10061-02-6trans-1, 3-Dichloropropene       16       U         75-25-2Bromoform       16       U         108-10-14-Methyl-2-Pentanone       16       U         591-78-62-Hexanone       16       U         127-18-4Totuene       16       U         108-88-31,1,2,2-Tetrachloroethane       16       U         108-88-3			16	U
67-66-3			16	U
78-93-32-Butanone       310       U         71-55-61,1,1-Trichloroethane       16       U         56-23-5Carbon Tetrachloride       16       U         75-27-4Bromodichloromethane       16       U         78-87-51,2-Dichloropropane       16       U         10061-01-5cis-1,3-Dichloropropene       16       U         79-01-6Trichloroethene       16       U         124-48-1Dibromochloromethane       16       U         79-00-51,1,2-Trichloroethane       16       U         71-43-2Benzene       16       U         1061-02-6trans-1,3-Dichloropropene       16       U         108-10-14-Methyl-2-Pentanone       16       U         127-18-62-Hexanone       16       U         127-18-4Tetrachloroethene       16       U         108-88-3Toluene       16       U         108-88-3Chlorobenzene       16       U         100-41-4Styrene       16       U	67-66-3	Chloroform	16	ש
78-93-32-Butanone       310       U         71-55-61,1,1-Trichloroethane       16       U         56-23-5Carbon Tetrachloride       16       U         75-27-4Bromodichloromethane       16       U         78-87-51,2-Dichloropropane       16       U         10061-01-5cis-1,3-Dichloropropene       16       U         79-01-6Trichloroethene       16       U         124-48-1Dibromochloromethane       16       U         79-00-51,1,2-Trichloroethane       16       U         71-43-2Benzene       16       U         1061-02-6trans-1,3-Dichloropropene       16       U         108-10-14-Methyl-2-Pentanone       16       U         127-18-62-Hexanone       16       U         127-18-4Tetrachloroethene       16       U         108-88-3Toluene       16       U         108-88-3Chlorobenzene       16       U         100-41-4Styrene       16       U	107-06-2	1,2-Dichloroethane	16	U
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78-87-51, 2-Dichloropropane       16       U         10061-01-5cis-1, 3-Dichloropropene       16       U         79-01-6Trichloroethene       16       U         124-48-1Dibromochloromethane       16       U         79-00-51, 1, 2-Trichloroethane       16       U         71-43-2Benzene       16       U         10061-02-6trans-1, 3-Dichloropropene       16       U         75-25-2Bromoform       16       U         108-10-14-Methyl-2-Pentanone       160       U         591-78-62-Hexanone       160       U         127-18-4Tetrachloroethene       16       U         79-34-51,1,2,2-Tetrachloroethane       16       U         108-88-3Toluene       16       U         108-90-7Chlorobenzene       16       U         100-41-4Styrene       16       U			16	ט
10061-01-5cis-1,3-Dichloropropene       16       U         79-01-6Trichloroethene       16       U         124-48-1Dibromochloromethane       16       U         79-00-51,1,2-Trichloroethane       16       U         71-43-2Benzene       16       U         10061-02-6trans-1,3-Dichloropropene       16       U         75-25-2Bromoform       16       U         108-10-14-Methyl-2-Pentanone       160       U         591-78-62-Hexanone       160       U         127-18-4Tetrachloroethene       16       U         79-34-51,1,2,2-Tetrachloroethane       16       U         108-88-3Toluene       16       U         108-90-7Chlorobenzene       16       U         100-41-4Styrene       16       U	75-27-4	Bromodichloromethane	16	ט
79-01-6Trichloroethene       16       U         124-48-1Dibromochloromethane       16       U         79-00-51,1,2-Trichloroethane       16       U         71-43-2Benzene       16       U         10061-02-6trans-1,3-Dichloropropene       16       U         75-25-2Bromoform       16       U         108-10-14-Methyl-2-Pentanone       160       U         591-78-62-Hexanone       160       U         127-18-4Tetrachloroethene       16       U         79-34-51,1,2,2-Tetrachloroethane       16       U         108-88-3Toluene       16       U         108-90-7Chlorobenzene       16       U         100-41-4Ethylbenzene       16       U         100-42-5Styrene       16       U	78-87-5	1,2-Dichloropropane	16	U
79-01-6Trichloroethene       16       U         124-48-1Dibromochloromethane       16       U         79-00-51,1,2-Trichloroethane       16       U         71-43-2Benzene       16       U         10061-02-6trans-1,3-Dichloropropene       16       U         75-25-2Bromoform       16       U         108-10-14-Methyl-2-Pentanone       160       U         591-78-62-Hexanone       160       U         127-18-4Tetrachloroethene       16       U         79-34-51,1,2,2-Tetrachloroethane       16       U         108-88-3Toluene       16       U         108-90-7Chlorobenzene       16       U         100-41-4Ethylbenzene       16       U         100-42-5Styrene       16       U	10061-01-5	cis-1,3-Dichloropropene	16	U
79-00-51,1,2-Trichloroethane       16       U         71-43-2Benzene       16       U         10061-02-6trans-1,3-Dichloropropene       16       U         75-25-2Bromoform       16       U         108-10-14-Methyl-2-Pentanone       160       U         591-78-62-Hexanone       160       U         127-18-4Tetrachloroethene       16       U         79-34-51,1,2,2-Tetrachloroethane       16       U         108-88-3Toluene       16       U         100-41-4Ethylbenzene       16       U         100-42-5Styrene       16       U	79-01-6	Trichloroethene	16	U
71-43-2Benzene       16       U         10061-02-6trans-1,3-Dichloropropene       16       U         75-25-2Bromoform       16       U         108-10-14-Methyl-2-Pentanone       160       U         591-78-62-Hexanone       160       U         127-18-4Tetrachloroethene       16       U         79-34-51,1,2,2-Tetrachloroethane       16       U         108-88-3Toluene       16       U         108-90-7Chlorobenzene       16       U         100-41-4Styrene       16       U	124-48-1	Dibromochloromethane	16	U
10061-02-6trans-1,3-Dichloropropene       16       U         75-25-2Bromoform       16       U         108-10-14-Methyl-2-Pentanone       160       U         591-78-62-Hexanone       160       U         127-18-4Tetrachloroethene       16       U         79-34-51,1,2,2-Tetrachloroethane       16       U         108-88-3Toluene       16       U         108-90-7Chlorobenzene       16       U         100-41-4Styrene       16       U	79-00-5	1,1,2-Trichloroethane	16	U
75-25-2Bromoform       16       U         108-10-14-Methyl-2-Pentanone       160       U         591-78-62-Hexanone       160       U         127-18-4Tetrachloroethene       16       U         79-34-51,1,2,2-Tetrachloroethane       16       U         108-88-3Toluene       16       U         108-90-7Chlorobenzene       16       U         100-41-4Styrene       16       U	71-43-2	Benzene	16	U
108-10-14-Methyl-2-Pentanone       160       U         591-78-62-Hexanone       160       U         127-18-4Tetrachloroethene       16       U         79-34-51,1,2,2-Tetrachloroethane       16       U         108-88-3Toluene       16       U         108-90-7Chlorobenzene       16       U         100-41-4Ethylbenzene       16       U         100-42-5Styrene       16       U	10061-02-6	trans-1,3-Dichloropropene	16	U
591-78-62-Hexanone       160       U         127-18-4Tetrachloroethene       16       U         79-34-51,1,2,2-Tetrachloroethane       16       U         108-88-3Toluene       16       U         108-90-7Chlorobenzene       16       U         100-41-4Ethylbenzene       16       U         100-42-5Styrene       16       U	75-25-2	Bromoform	16	ט
591-78-62-Hexanone       160       U         127-18-4Tetrachloroethene       16       U         79-34-51,1,2,2-Tetrachloroethane       16       U         108-88-3Toluene       16       U         108-90-7Chlorobenzene       16       U         100-41-4Ethylbenzene       16       U         100-42-5Styrene       16       U	108-10-1	4-Methyl-2-Pentanone	160	ט
79-34-51,1,2,2-Tetrachloroethane       16       U         108-88-3Toluene       16       U         108-90-7Chlorobenzene       16       U         100-41-4Ethylbenzene       16       U         100-42-5Styrene       16       U			160	U
108-88-3Toluene       16       U         108-90-7Chlorobenzene       16       U         100-41-4Ethylbenzene       16       U         100-42-5Styrene       16       U	127-18-4	Tetrachloroethene	16	U
108-88-3Toluene       16       U         108-90-7Chlorobenzene       16       U         100-41-4Ethylbenzene       16       U         100-42-5Styrene       16       U	79-34-5	1,1,2,2-Tetrachloroethane	16	U
100-41-4Ethylbenzene 16 U 100-42-5Styrene 16 U			16	Ū
100-41-4Ethylbenzene 16 U 100-42-5Styrene 16 U	108-90-7	Chlorobenzene	16	U
100-42-5Styrene16 U			16	[ט
1330-20-7Xylene (total) 16 U			16	U
			16	U

3/27/95 5

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#### 1A VOLATILE ORGANICS ANALYSIS DATA SHEE

EPA SAMPLE NO. BHI-00405

Rev. 00 BODMT4

Lab Name: QUANTERRA MO

Contract: 550-56

Lab Code: ITMP Case No.: V44408

SDG No.: W0430

Matrix: (soil/water) SOIL

Lab Sample ID: 7444-008

Sample wt/vol:

5.00 (g/mL) G

Lab File ID: E2000

Level: (low/med) LOW

Date Received: 01/30/95

% Moisture: not dec. 6

Date Analyzed: 02/06/95

Column: (pack/cap) CAP

Dilution Factor: 1.0

CONCENTRATION UNITS:

CAS NO.

COMPOUND

(ug/L or ug/Kg) UG/KG Q

74-87-3	Chloromethane	11	U
	Bromomethane	11	טן
	Vinyl Chloride	11	ן ט -
	Chloroethane	11.	U .
	Methylene Chloride	11 1	B/ M
67-64-1		110	ับ
75-15-0	Carbon Disulfide	5	U
75-35-4 <b></b> -	1,1-Dichloroethene	5	U
	1,1-Dichloroethane	5	ט
540-59-0	1,2-Dichloroethene (total)	5	U
67-66-3	Chloroform	5	U
107-06-2	1,2-Dichloroethane	5	U
78-93-3- <del></del> -		110	U
71-55-6	1,1,1-Trichloroethane	5	U
56-23-5	Carbon Tetrachloride	5	U
75-27-4	Bromodichloromethane	5	U
	1,2-Dichloropropane	5	שׁ
	cis-1,3-Dichloropropene	5	U
	Trichloroethene	5	ט
<del></del>	Dibromochloromethane	5	U
79-00-5 <b></b> -	1,1,2-Trichloroethane	5	U
71-43-2		5	U
10061-02-6	trans-1,3-Dichloropropene	5	ט
75-25-2		5	ן ט
	4-Methyl-2-Pentanone	53	U
	2-Hexanone	53	U
	Tetrachloroethene	5	U
	1,1,2,2-Tetrachloroethane	5	U
108-88-3 <del>-</del> -		5	U
	Chlorobenzene	5	U
	Ethylbenzene	5	U
100-42-5		5	U
1330-20-7	Xylene (total)	5	U

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# APPENDIX C SEMIVOLATILE ORGANIC DATA SUMMARY TABLES

				_																	
Project: WESTINGHOUSE-HANF	ORD			]																	
Laboratory: QUANTERRA				]																	
Case:	SDG:	W0429		<u> </u>																	
Sample Number		BODMT2		BODMT3														<u> </u>			
Location		Test Pit #	<b>#2</b>	Test Pit	#2_																
Remarks																L		l			
Sample Date		01/17/95		01/17/95	5									<u> </u>							
Extraction Date		01/20/95		01/20/95																	
Analysis Date		01/23/95		01/23/95	<u> </u>			<u> </u>				<u> </u>			<u> </u>			<u></u>		<u> </u>	<del></del> -
				Result		Result	Q	Result	Q	Result	Q	Result	Q	Result	a	Result	Q	Result	Q	Result	1Q
Phenol	330	1200		2100	ĺΠ		<u> </u>		<u> </u>	<u> </u>	<u> </u>	<u> </u>	<u> </u>	<u> </u>		<u> </u>	<u> </u>	<u> </u>	<u> </u>		1
bis(2-Chloroethyl)ether	330	1200		2100	Ĭ'n	<u> </u>	<u> </u>	<u> </u>	<u> </u>		<b> </b>				<b> </b>	<b></b> _	<u> </u>	<u> </u>	ऻ		<del> </del>
2-Chiorophenol	330	1200	ļΨ	2100		<b> </b>	L_	<b> </b>	<b> </b>	ļ	<b> </b>	<u> </u>	<u> </u>	<b> </b> _	<u> </u>	<b> </b>	Щ.	ļ	<del> </del>	<u> </u>	4—
1,3-Dichlorobenzene	330	1200		2100	밥	<u> </u>	<b>!</b> —	<u> </u>	<b>!</b>		<del> </del>	<u> </u>	<u> </u>	<u> </u>		ļ	<b>L</b>	<b> </b> _	ـــــ	ļ	┸
1,4-Dichlorobenzene	330	1200		2100	빈	<b></b>	<u> </u>	<b> </b>	┡-	<b> </b> _	<del> </del> _	<b> </b>	L_	ļ	1	<b> </b>	<u> </u>		<b>!</b>		4
1,2-Dichlorobenzene	330	1200		2100		<u> </u>	L	<u> </u>			┖				<u> </u>	ļ	<u> </u>		1	<u> </u>	
2-Methylphenol	330	1200		2100		L	<u> </u>	L	<u> </u>	l	<u> </u>	L	<u> </u>	<u> </u>	<b>I</b>	<u> </u>	1_		<u> </u>	<u> </u>	
2,2'-oxybis(1-Chloropropane)	330	1200		2100		<u> </u>		<u> </u>	L		<u> </u>			<u> </u>	<u> </u>		L	<u> </u>	<u> </u>	<u> </u>	
4-Methylpheno	330	1200		2100	U	<u> </u>	<u> </u>				1				L			<u> </u>		<u> </u>	1_
N-Nitroso-di-n-propylamine	330	1200		2100	טן				$\Gamma$	<u> </u>	$\Gamma$		L		$\Box$				L	I	$T_{-}$
Hexachloroethane	330	1200		2100							Г		1				T		П		T
Nitrobenzene	330	1200		2100	U										$\Box$				T		T
Isophorone	330	1200	U	2100	U		T -				1	T					T				_
2-Nitrophenol	330	1200		2100	U		1		T		T		Г					<u> </u>	o	1	T
2,4-Dimethylphenol	330	1200	U	2100	U	i	1-	1	1		1								$\top$	1	1
bis(2-Chloroethoxy)methane	330	1200	U	2100	U		1		1		1			<u> </u>			1		1		1
2,4-Dichlorophenol	330	1200	U	2100	U	1								i							1
1,2,4-Trichlorobenzene	330	1200	U	2100	<u>ַ</u>	<del>                                     </del>	<u> </u>		1		<b>T</b>		<b></b>				1	1	T	<del>                                     </del>	_
Naphthalene	330	1200	U	2100	U	1	1	<del></del>	1	1	1		1				T-	<u> </u>	1-		
4-Chloroaniline	1700	2300		4100		T	Τ		Т		$\top$	T	Π	1		<u> </u>	1	1	$\top$		T
Hexachlorobutadiene	330	1200		2100		1	1	T	1	Γ	1	]	<u> </u>	]	T_	I — —	1	1	1	<del>,                                      </del>	1
4-Chloro-3-methylphenol	1700	2300		4100	U	<u> </u>	Τ		Π				T			<u> </u>	T	T	$\top$		<del>                                      </del>
2-Methylnaphthalene	330	1200		2100	U	I			Π	<u> </u>		1	Γ	<u> </u>	T		Τ	Ī	1	1	T
Hexachlorocyclopentadiene	330	1200	U	2100		T	1				T	]	$\Gamma$		$I^{-}$	1	$\Gamma$		7		$\top$
2,4,6-Trichiorophenol	330	1200	U	2100			T	T	$I^{-}$		T	<u> </u>	T		T	1	T_	1	1	T	$\top$
2,4,5-Trichlorophenol	330	1200		2100	U	1	ī		T		$\top$		Π	<u> </u>	Γ	· ·	Т	1	T	<del> </del>	1
2-Chloronaphthalene	330	1200	U	2100		1	1		1	1	1		1		$\Gamma$		1	1	1		1
2-Nitroaniline	1700	5900		10000		<b>T</b>			1		T		<u> </u>			<del>   </del>	1		1		1
Dimethyl phthalate	330	1200		2100		1	1	<del> </del>	1		1	1	T	<u> </u>		<del>                                     </del>	1	1	†	<del> </del>	1
Acenaphthylene	330	1200		2100		1	1	<del>                                     </del>	1		$\top$	1	1	<del>                                     </del>	1			1	1		$\top$
2,6-Dinitrotoluene	330	1200		2100		<del>                                     </del>	Τ_	<del></del>	1		1	1	T		1	<del>                                     </del>	1	<del> </del>	1	<del> </del>	7

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Project: WESTINGHOUSE-HANF	-OHD			]																	
Laboratory: QUANTERRA																					
		W0430		ļ				,		<del></del>											
Sample Number		BODMT4		<u> </u>		<u> </u>		<u> </u>		<b> </b>						ļ		<u> </u>			
Location		Test Pit a	#2	<u> </u>		Ĺ. <u>.</u>				<u> </u>		Í		<b> </b>		Ĺ <u> </u>		ļ			
Remarks				<u> </u>				<u> </u>				<u> </u>		<u> </u>		<b> </b>		<u> </u>			
Sample Date		01/27/95	5					<u> </u>		<u> </u>				L		<u> </u>		<u> </u>			
Extraction Date		02/02/95	<u> </u>	ļ										<u> </u>				<b> </b>			
Analysis Date		02/06/95		<u> </u>				<u> </u>	-	ļ			-		1 4		7.8		1.8	B	78
				Result	Q	Result	Q	Result	<u>ja</u>	Result	Q	Hesut	10								
Phenol	330	700			l	L	1		L.	<u></u>	<u> </u>		<u> </u>	L	<u> </u>	<u> </u>	<u>L.</u>	<u> </u>	<u> </u>	<u> </u>	<u></u>
bis(2-Chloroethyl)ether	330	700			П			[			Γ	L	<u> </u>			Ĺ	L.,				<u> </u>
2-Chlorophenol	330				Г		П	Γ	П	-		I	Γ		}	I			$\Gamma$		
1,3-Dichlorobenzene	330	700			1		Π	1	Т		Γ	<u> </u>	Г		ĺ		Π				
1,4-Dichlorobenzene	330	700		1	T	1	Г		Τ		<b>1</b>			T	Ī	<u> </u>			Π		Γ
1,2-Dichlorobenzene	330	700	U	<u> </u>	†	<del> </del>	1	† — — — — — — — — — — — — — — — — — — —	1		i							1			$\vdash$
2-Methylphenol	330	700		1	1-		l	<del>                                     </del>	╁	i		<del>                                     </del>				<u> </u>	1	1	1		一
2,2'-oxybis(1-Chloropropane)	330			<b></b>	1	<del> </del>	1	<del> </del>	<del>                                     </del>	<del>                                     </del>	<del>                                     </del>	<del>                                     </del>	<del>                                     </del>	<del>                                     </del>	1-	<del></del>	1	<del> </del>	1		† <del></del>
4-Methylphenol	330	700		<del></del>	1		f—	<del></del>	一	<del> </del>	_			f	<del> </del>	t	<del>  -</del>	<del>                                     </del>	1-	<del></del>	+-
N-Nitroso-di-n-propylamine	330			<del> </del>	╁	<del> </del>	<del> </del>	<del> </del>	╂-		┨	<del> </del> -	<del>                                     </del>		t	ļ — — —	╅╌	<del> </del>	<del> </del>	<del> </del>	+
Hexachloroethane	330	700		<del> </del>	╁╾	<del> </del>	<del>                                     </del>	<del> </del>	┼	<del> </del>	<del>                                     </del>	<del>}</del> -			╀─						
Nitrobenzene	330	700		<del> </del>	╁	<del> </del>	1-	<del> </del> -	╁	<del> </del>	┢	<del> </del> -	┈	<del> </del> -	┼─	<del> </del>	╁╌	<del> </del>	╁	<del> </del> -	+
Isophorone	330			<del> </del>		ļ	<del> </del>	<del> </del>	╂	<del> </del> -	<del>                                     </del>	<del> </del>	<del> </del> —	<del> </del>	<del>                                     </del>	<del> </del>	┼	<del>├</del> -	<del> </del>	<del> </del> -	┯
2-Nitrophenol	330			<del> </del>	<del> </del>	<del> </del> -	<b>├</b> ──	<del> </del> -	┼	<del> </del>	<del> </del> —	<del> </del>	<del> </del> —	<del>}</del>	╀	<del> </del>	<del> </del>	<del> </del>	<del> </del>	<del> </del> -	╀
	330			<del> </del>	1-		—	<del> </del>	1-	<del> </del>	<del> </del> —	<del> </del>	<del>  -</del>		╌	<del> </del> -	<b>↓</b> —	<del> </del> -	ļ	<del></del>	┾-
2,4-Dimethylphenol				<del> </del>	<del>↓</del> —	<del> -</del>	₩	<del> </del>	<del> </del>	<del> </del>	<u> </u>	ļ	<u> </u>	ļ	ļ	<del> </del>	1-	<del> </del>	┼—	<del> </del>	┼
bis(2-Chloroethoxy) methane	330			<del> </del>	1-	ļ	<del> </del>	<del> </del>	1-	<del> </del> -	ļ	<del> </del>	<del> </del>	<b>Ļ</b>	<b>↓</b> —	ļ	<del> </del>	<del>                                     </del>	ـ	ļ	∔—
2,4-Dichlorophenol	330			ļ	<b>.</b>	ļ	↓	<u> </u>	<del> </del>	<del> </del>	<del>  _</del>	ļ	<u> </u>	ļ	1	ļ	4—	<b>↓</b>	<b> </b>	<b> </b>	Д—
1,2,4-Trichlorobenzene	330			<b></b> _	ļ	<u> </u>	<u> </u>	ļ	<b>Ļ</b>	<b></b>	<u> </u>	ļ	<u> </u>	\	<b>_</b>	Ļ	<del> </del> _	<u> </u>	↓_		—
Naphthalene	330			<u> </u>		<u> </u>	<u> </u>	<u> </u>	<u> </u>	↓	<u> </u>	<u> </u>	L	<u> </u>	╙	<u> </u>	1_				
4-Chloroaniline	1700			<u> </u>		<u> </u>	ļ	<u> </u>				1	<u> </u>			<u> </u>	<u> </u>		1_	<u> </u>	L
Hexachlorobutadiene	330						1	L'	_						<u> </u>	<u></u>	<u> </u>	J	<u> </u>		L
4 Chloro3 methylphenol	1700								Ι		Ι.						1				
2-Methylnaphthalene	330								Π.												
Hexachiorocyclopentadiene	330			T	1	<u> </u>	<b>T</b>	<del>                                     </del>	$\Box$		Г		<b>!</b>		$\top$		1				$\sqcap$
2,4,6-Trichlorophenol	330	700	U		1	T	T	1	1-	1	1		Т		1				T		1
2,4,5-Trichlorophenol	330	700	ΙŪ		1	†	1	<del>                                     </del>	$\top$	1	1	1	1	<del></del>	Τ	1	1	<del></del>		1	$\top$
2-Chloronaphthalene	330			1	1	<del>                                     </del>	1	1	1	1	1		1	1	T	1	7	<del>                                     </del>	T	1	T
2-Nitroaniline	1700		lŪ		$\top$	T		1	1	<del>                                     </del>	T	<del>                                     </del>	t	†			1	T	1		1
Dimethyl phthalate	330			<del> </del> -	<del>                                     </del>	<del>                                     </del>	+	<del> </del>	1	<del>                                     </del>	T	<del> </del>	1	t	十一	<del> </del>	1	† — —	<del>                                     </del>	<del>                                     </del>	$\top$
Acenaphthylene	330			┼───	╁┈	<del>                                     </del>	1	<del> </del>	1—	<del> </del>	$\vdash$	<del> </del>	-	<del> </del>	1	<del> </del>	$t^-$	<del> </del>	<del> </del>	<del>                                     </del>	+-
2,6-Dinitrotoluene	330			<del> </del> -	+-	<del> </del>	+	<del> </del> -	+	<del> </del>	┰	<del> </del>	├─	<del> </del>	<del> </del>	<del>                                     </del>	1-	<del> </del>	+-	<del> </del> -	7   V
Lio Milliordiscito		1	T~_	<del></del> _	1	<del></del>	Ь.	<b>↓</b>		<del></del>	┺	<del></del>						.L	ــــــــــــــــــــــــــــــــــــــ	V25	4-6

### APPENDIX D

### SEMIVOLATILE ORGANIC VALIDATED LABORATORY REPORT FORMS

#### 1B SEMIVOLATILE ORGANICS ANALYSIS DATA SHEET

EPA SAMPLE NO BHI-00405

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Rev. 00

BODMT2

Lab Name: OUANTERRA MO Contract: 550-56

COMPOUND

CAS NO.

108-95-2----Phenol

Lab Code: ITMO Case No.: S34402 SAS No.: SDG No.: W0429

Matrix: (soil/water) SOIL Lab Sample ID: 7344-002

Sample wt/vol: 30.00 (g/mL) G Lab File ID: A7716

Level: (low/med) LOW Date Received: 01/18/95

% Moisture: not dec. 44 dec. Date Extracted: 01/20/95

Extraction: (SepF/Cont/Sonc) SONC Date Analyzed: 01/23/95

GPC Cleanup: (Y/N) N Dilution Factor: 1.00 pH:

CONCENTRATION UNITS:

(ug/L or ug/Kg) UG/KG

111-44-4-----bis(2-Chloroethyl)Ether 1200 U 95-57-8-----2-Chlorophenol 1200 U 541-73-1----1,3-Dichlorobenzene\_ 1200 U 106-46-7----1, 4-Dichlorobenzene 1200 U 95-50-1----1,2-Dichlorobenzene U 1200 95-48-7----2-Methylphenol 1200 Ŭ 108-60-1----2,2'-oxybis(1-Chloropropane) U 1200 106-44-5-----4-Methylphenol 1200 U 621-64-7----N-Nitroso-Di-n-Propylamine 1200 U 67-72-1-----Hexachloroethane Ū 1200 98-95-3-----Nitrobenzene 1200 U 78-59-1-----Isophorone U 1200 88-75-5----2-Nitrophenol 1200 U 105-67-9-----2,4-Dimethylphenol U 1200 111-91-1-----bis(2-Chloroethoxy)Methane U 1200 120-83-2----2,4-Dichlorophenol 1200 U 120-82-1----1,2,4-Trichlorobenzene 1200 U

3/27/95 50

91-20-3-----Naphthalene

106-47-8-----4-Chloroaniline

88-74-4----2-Nitroaniline

208-96-8-----Acenaphthylene

87-68-3-----Hexachlorobutadiene

91-57-6----2-Methylnaphthalene

88-06-2----2,4,6-Trichlorophenol

95-95-4----2,4,5-Trichlorophenol\_

91-58-7----2-Chloronaphthalene

131-11-3-----Dimethyl Phthalate

606-20-2----2,6-Dinitrotoluene

59-50-7----4-Chloro-3-Methylphenol

77-47-4------Hexachlorocyclopentadiene

FORM I SV-1

SEMIVOLATILE ORGANICS ANALYSIS DATA SHEET TENTATIVELY IDENTIFIED COMPOUNDS

BHI-00405 Rev. 00 B0DMT2

EPA SAMPLE NO.

Lab Name: QUANTERRA MO

Contract: 550-56

Lab Code: ITMO Case No.: S34402 SAS No.:

SDG No.: W0429

Matrix: (soil/water) SOIL

Lab Sample ID:

7344-002

Sample wt/vol:

30.00 (q/mL) G

Lab File ID:

A7716

Level: (low/med) LOW

Date Received: 01/18/95

% Moisture: not dec. 44

dec.

Date Extracted: 01/20/95

Extraction: (SepF/Cont/Sonc) SONC

Date Analyzed: 01/23/95

GPC Cleanup: (Y/N) N

pH:

Dilution Factor: 1.00

Number TICs found: 22

CONCENTRATION UNITS: (ug/L or ug/Kg) UG/KG

CAS NUMBER	COMPOUND NAME	RT	EST. CONC.	Q
1. 0	Aldol Condensation	4.64	25000	ABJ U
2.	Aldol Condensation	4.70	2600	ABJU
3.	UNKNOWN	16.77	240	J
4.	UNKNOWN	18.59	2800	J
5.	Unknown Alkane	18.84	1200	J
6.	Unknown Alkane	18.94	320	ĺJ
7.	Unknown Alkane	19.44	290	J
8.	UNKNOWN	20.50	390	J
9.	Unknown Cl2H6Cl4	22.92	220	J
10.	Unknown Cl2H6Cl4	23.02	340	J
11.	Unknown Cl2H5Cl5	23.53	1400	J
12.	Unknown Cl2H5Cl5	24.07	590	J
13.	Unknown C12H4Cl6	24.46	560	{J
14.	Unknown C12H4Cl6	24.70	990	J
15.	Unknown Cl2H5Cl5	24.79	900	J
16.	Unknown C12H4Cl6	25.22	1100	J
17.	Unknown Cl2H4Cl6	25.75	1200	J
18.	Unknown C12H3Cl7	26.02	500	J
19.	Unknown C12H3Cl7	27.03	650	J
20.	UNKNOWN	28.32	610	J
21.	UNKNOWN	29.60	1000	J
22.	UNKNOWN	29.88	1100	J
		<u></u>		

3/27/955

# SEMIVOLATILE ORGANICS ANALYSIS DATA SHEET

EPA SAMPLE NO. BHI-00405

Rev. 00 BODMT3

Lab Name: QUANTERRA MO Contract: 550-56

Lab Code: ITMO Case No.: S34402 SAS No.: SDG No.: W0429

Matrix: (soil/water) SOIL Lab Sample ID: 7344-004

Sample wt/vol: 30.00 (g/mL) G Lab File ID: A7718

Level: (low/med) LOW Date Received: 01/18/95

% Moisture: not dec. 68 dec. Date Extracted: 01/20/95

Extraction: (SepF/Cont/Sonc) SONC Date Analyzed: 01/23/95

GPC Cleanup: (Y/N) N pH: Dilution Factor: 1.00

CONCENTRATION UNITS:

CAS NO.	COMPOUND (ug/L or ug)	/Kg) UG/KG	Q	
99-09-2	3-Nitroaniline	10000	טו	
	Acenaphthene	2100	υ -	
	2,4-Dinitrophenol	10000	U	- {
	4-Nitrophenol_	10000	Ū	
	Dibenzofuran	2100	ប	ĺ
121-14-2	2,4-Dinitrotoluene	2100	ט	}
	Diethylphthalate	2100	ט	
7005-72-3	4-Chlorophenyl-phenylether	2100	U	
86-73-7	Fluorene	2100	ט	
	4-Nitroaniline	4100	Ū	
	4,6-Dinitro-2-Methylphenol	10000	ָ <sup>±</sup>	
	N-Nitrosodiphenylamine (1)	2100	שׁ	
101-55-3	4-Bromophenyl-phenylether	2100	ט	-
	Hexachlorobenzene	2100	Ū	
	Pentachlorophenol	10000	Ū	1
85-01-8	Phenanthrene	470	J	
120-12-7	Anthracene	460	Ĵ	
	Carbazole	2100	ĺυ	ĺ
84-74-2	Di-n-Butylphthalate	2100	U	
206-44-0	Fluoranthene	940	J	
129-00-0	Pyrene	820	J	- {
	Butylbenzylphthalate	2100 1400	BU	- I
91-94-1	3,3'-Dichlorobenzidine	4100	Ü	1
56-55-3	Benzo(a)Anthracene	340	IJ	-
	Chrysene	400	J	
117-81-7	bis(2-Ethylhexyl)Phthalate	130	J	1
117-84-0	Di-n-Octyl Phthalate	2100	M	Į,
205-99-2	Benzo(b)Fluoranthene	580	7	
	Benzo(k)Fluoranthene	220	Ø	
	Benzo(a) Pyrene	290	5	
193-39-5	Indeno(1,2,3-cd)Pyrene	2100	7	
53-70-3	Dibenz (a, h) Anthracene	2100	ช	
191-24-2	Benzo(g,h,i)Perylene	2100	Ø	ı
) - Cannot h	oe separated from Diphenylamine	I	_	‡

#### SEMIVOLATILE ORGANICS ANALYSIS DATA SHEET

BHI-00405 Rev. 00 BODMT4

Q

Lab Name: QUANTERRA MO Contract: 550-56

Lab Code: ITMO Case No.: S44408 SAS No.: SDG No.: W0430

Matrix: (soil/water) SOIL Lab Sample ID: 7444-008

Sample wt/vol: 30.00 (g/mL) G Lab File ID: D6748

Level: (low/med) LOW Date Received: 01/30/95

% Moisture: not dec. 6 dec. Date Extracted: 02/02/95

Extraction: (SepF/Cont/Sonc) SONC Date Analyzed: 02/06/95

GPC Cleanup: (Y/N) N pH: Dilution Factor: 1.00

CAS NO. COMPOUND CONCENTRATION UNITS: (ug/L or ug/Kg) UG/KG

108-95-2Phenol		<del></del>	
111-44-4	(08-95-2Pheπol	700	U
95-57-82-Chlorophenol       700       U         541-73-11,3-Dichlorobenzene       700       U         106-46-71,4-Dichlorobenzene       700       U         95-50-11,2-Dichlorobenzene       700       U         95-48-72-Methylphenol       700       U         108-60-12,2'-oxybis (1-Chloropropane)       700       U         106-44-54-Methylphenol       700       U         621-64-7Nitroso-Di-n-Propylamine       700       U         67-72-1Hexachloroethane       700       U         98-95-3Nitrobenzene       700       U         78-59-1		<u> </u>	1 -
541-73-11,3-Dichlorobenzene       700       U         106-46-71,4-Dichlorobenzene       700       U         95-50-11,2-Dichlorobenzene       700       U         95-48-72-Methylphenol       700       U         108-60-12,2'-oxybis (1-Chloropropane)       700       U         106-44-54-Methylphenol       700       U         621-64-7Nnitroso-Di-n-Propylamine       700       U         67-72-1Hexachloroethane       700       U         98-95-3Nitrobenzene       700       U         88-75-52-Nitrophenol       700       U         105-67-92,4-Dimethylphenol       700       U         111-91-1bis (2-Chloroethoxy) Methane       700       U         120-83-22,4-Dichlorophenol       700       U         120-82-11,2,4-Trichlorobenzene       700       U         91-20-3Naphthalene       700       U         106-47-8			1 -
106-46-71,4-Dichlorobenzene		<b>  </b>	I -
95-50-11, 2-Dichlorobenzene       700       U         95-48-72-Methylphenol       700       U         108-60-12, 2'-oxybis (1-Chloropropane)       700       U         106-44-54-Methylphenol       700       U         621-64-7N-Nitroso-Di-n-Propylamine       700       U         67-72-1Hexachloroethane       700       U         98-95-3Nitrobenzene       700       U         78-59-1Isophorone       700       U         88-75-52-Nitrophenol       700       U         105-67-92,4-Dimethylphenol       700       U         111-91-1bis (2-Chloroethoxy) Methane       700       U         120-83-22,4-Dichlorophenol       700       U         120-82-11,2,4-Trichlorobenzene       700       U         91-20-3Naphthalene       700       U         106-47-8		_1	_
95-48-72-Methylphenol       700       U         108-60-12,2'-oxybis (1-Chloropropane)       700       U         106-44-54-Methylphenol       700       U         621-64-7Nitroso-Di-n-Propylamine       700       U         67-72-1Hexachloroethane       700       U         98-95-3Nitrobenzene       700       U         78-59-1		<b></b>	1 -
108-60-12,2'-oxybis(1-Chloropropane)       700       U         106-44-54-Methylphenol       700       U         621-64-7Nhitroso-Di-n-Propylamine       700       U         67-72-1Hexachloroethane       700       U         98-95-3Nitrobenzene       700       U         78-59-1Isophorone       700       U         88-75-52-Nitrophenol       700       U         105-67-92,4-Dimethylphenol       700       U         111-91-1bis(2-Chloroethoxy)Methane       700       U         120-83-22,4-Dichlorophenol       700       U         120-82-11,2,4-Trichlorobenzene       700       U         91-20-3Naphthalene       700       U         106-47-8		<b>-</b> 1	_
106-44-54-Methylphenol       700       U         621-64-7N-Nitroso-Di-n-Propylamine       700       U         67-72-1Hexachloroethane       700       U         98-95-3Nitrobenzene       700       U         78-59-1Isophorone       700       U         88-75-52-Nitrophenol       700       U         105-67-92,4-Dimethylphenol       700       U         111-91-1bis(2-Chloroethoxy)Methane       700       U         120-83-22,4-Dichlorophenol       700       U         120-82-11,2,4-Trichlorobenzene       700       U         91-20-3Naphthalene       700       U         106-47-8			I -
621-64-7N-Nitroso-Di-n-Propylamine       700       U         67-72-1Hexachloroethane       700       U         98-95-3Nitrobenzene       700       U         78-59-1Isophorone       700       U         88-75-52-Nitrophenol       700       U         105-67-92,4-Dimethylphenol       700       U         111-91-1bis (2-Chloroethoxy) Methane       700       U         120-83-22,4-Dichlorophenol       700       U         120-82-11,2,4-Trichlorobenzene       700       U         91-20-3Naphthalene       700       U         106-47-8Naphthalene       700       U         87-68-3		<del>-</del> I	
67-72-1		<del></del> (	1 -
98-95-3Nitrobenzene       700       U         78-59-1Isophorone       700       U         88-75-52-Nitrophenol       700       U         105-67-92,4-Dimethylphenol       700       U         111-91-1bis(2-Chloroethoxy)Methane       700       U         120-83-22,4-Dichlorophenol       700       U         120-82-11,2,4-Trichlorobenzene       700       U         91-20-3Naphthalene       700       U         106-47-8Naphthalene       700       U         87-68-3		<b></b> ]	, -
78-59-1		_	-
88-75-52-Nitrophenol       700       U         105-67-92,4-Dimethylphenol       700       U         111-91-1bis(2-Chloroethoxy)Methane       700       U         120-83-22,4-Dichlorophenol       700       U         120-82-11,2,4-Trichlorobenzene       700       U         91-20-3Naphthalene       700       U         106-47-8Naphthalene       700       U         87-68-3		<del> }</del>	J
105-67-92,4-Dimethylphenol       700       U         111-91-1bis (2-Chloroethoxy) Methane       700       U         120-83-22,4-Dichlorophenol       700       U         120-82-11,2,4-Trichlorobenzene       700       U         91-20-3Naphthalene       700       U         106-47-8		<b></b> 1	1 -
111-91-1bis (2-Chloroethoxy) Methane       700       U         120-83-22, 4-Dichlorophenol       700       U         120-82-11, 2, 4-Trichlorobenzene       700       U         91-20-3Naphthalene       700       U         106-47-8		<del>-</del> {	•
120-83-22,4-Dichlorophenol       700       U         120-82-11,2,4-Trichlorobenzene       700       U         91-20-3Naphthalene       700       U         106-47-84-Chloroaniline       1400       U         87-68-3Hexachlorobutadiene       700       U         59-50-74-Chloro-3-Methylphenol       1400       U         91-57-64-Chloro-3-Methylphenol       700       U         77-47-4	11-91-1his (2-Chloroethory) Methane		_
120-82-11,2,4-Trichlorobenzene       700       U         91-20-3Naphthalene       700       U         106-47-84-Chloroaniline       1400       U         87-68-3Hexachlorobutadiene       700       U         59-50-74-Chloro-3-Methylphenol       1400       U         91-57-62-Methylnaphthalene       700       U         77-47-4			1
91-20-3Naphthalene       700       U         106-47-84-Chloroaniline       1400       U         87-68-3Hexachlorobutadiene       700       U         59-50-74-Chloro-3-Methylphenol       1400       U         91-57-62-Methylnaphthalene       700       U         77-47-4		<del></del> {	1
106-47-84-Chloroaniline       1400       U         87-68-3Hexachlorobutadiene       700       U         59-50-74-Chloro-3-Methylphenol       1400       U         91-57-62-Methylnaphthalene       700       U         77-47-4Hexachlorocyclopentadiene       700       U         88-06-22,4,6-Trichlorophenol       700       U         95-95-42,4,5-Trichlorophenol       700       U		-1	1 -
87-68-3Hexachlorobutadiene       700       U         59-50-74-Chloro-3-Methylphenol       1400       U         91-57-62-Methylnaphthalene       700       U         77-47-4		I	i -
59-50-74-Chloro-3-Methylphenol       1400       U         91-57-62-Methylnaphthalene       700       U         77-47-4Hexachlorocyclopentadiene       700       U         88-06-22,4,6-Trichlorophenol       700       U         95-95-42,4,5-Trichlorophenol       700       U		- I	1 -
91-57-62-Methylnaphthalene       700       U         77-47-4Hexachlorocyclopentadiene       700       U         88-06-22,4,6-Trichlorophenol       700       U         95-95-42,4,5-Trichlorophenol       700       U		<b> l</b>	1 -
77-47-4Hexachlorocyclopentadiene 700 U 88-06-22,4,6-Trichlorophenol 700 U 95-95-42,4,5-Trichlorophenol 700 U		<b>-</b> ∤	] -
88-06-22,4,6-Trichlorophenol 700 U 95-95-42,4,5-Trichlorophenol 700 U		<b>-</b>	1 -
95-95-42,4,5-Trichlorophenol 700 U		<b>-</b> i	
		<del>-</del> 1	1 -
	91-58-72-Chloronaphthalene	-  700 700	U
88-74-42-Nitroaniline 3500 U		-1	-
131-11-3Dimethyl Phthalate 700 U		<u>-  </u>	] ~
208-96-8Acenaphthylene 700 U		)	} =
606-20-22,6-Dinitrotoluene 700 U		<b>-</b>	_
700	z, o panaga o o cuono	-	

3/20195 5

### SEMIVOLATILE ORGANICS ANALYSIS DATA SHEET TENTATIVELY IDENTIFIED COMPOUNDS

Rev. 00 BODMT4

BHI-00405

EPA SAMPLE N.

Contract: 550-56

Lab Code: ITMO Case No.: S44408 SAS No.: SDG No.: W0430

Matrix: (soil/water) SOIL

Lab Name: QUANTERRA MO

Lab Sample ID: 7444-008

Sample wt/vol: 30.00 (g/mL) G

Lab File ID: D6748

Level: (low/med) LOW

Date Received: 01/30/95

% Moisture: not dec. 6 dec.

Date Extracted: 02/02/95

Extraction: (SepF/Cont/Sonc) SONC Date Analyzed: 02/06/95

GPC Cleanup: (Y/N) N pH:

Dilution Factor: 1.00

CONCENTRATION UNITS: (ug/L or ug/Kg) UG/KG

Number TICs found: 2

CAS NUMBER	COMPOUND NAME	RT	EST. CONC.	Q =====
1. 0	Aldol Condensation UNKNOWN	5.31 29.13	16000 77	ABJ U

3/30/955

# APPENDIX E PESTICIDE/PCB DATA SUMMARY TABLES

	SDG:	W0429																<del></del>			
Sample Number		B0DMT2		BODMT3																	
Location		Test Pit #	#2	Test Pit ≠	F2																
Remarks																					
Sample Date		01/17/95		01/17/95																	
Extraction Date		01/20/95		01/20/95														ļ		ļ	
Analysis Date		01/25/95		01/25/95				1			-					ļ <u></u>	12		T =	<u> </u>	т=
Pesticide/PCB	CRQL			Result		Result	Q	Result	Q	Result	Q	Result	Q	Result	a	Result	Q	Result	Q	Result	Q
alpha - BHC	1.7		U	3.1		<u> </u>			<u> </u>		<u> </u>				<u> </u>	ļ	L		<b> </b>	<b> </b>	↓_
beta – BHC	1.7	3.5		6.2			L	L		<u> </u>					<u> </u>		<b>↓</b>	L	ـــــ	ļ	↓_
delta – BHC	1.7			9.3		l	<u> </u>	<u> </u>	<u> </u>		<u> </u>		$ldsymbol{ld}}}}}}$		<u> </u>		<u> </u>	<u> </u>	<u> </u>	ļ <u>.</u> .	<u> </u>
gamma-BHC (Lindan	) 1.7	4.5		4.1				l	<u> </u>		<u> </u>						<u> </u>		<b>L</b>		_
Heptachlor	1.7	1.8	UJ	3.1					<u> </u>				L				<b> </b>		<u> </u>		L.
Aldrin	1.7	8.2	J	36	J														<u> </u>		
Heptachlor epoxide	1.7		UJ	86	UJ		T														
Endosulfan i	1.7				U		П						I .								
Dieldrin	3.3		UJ	2.1	ÜJ		1	1		1	1								L		Γ
4,4'-DDE	3.3	25	ŪJ	82	IJ	1	1				1		<u> </u>	Ī							
Endrin	3.3			440	ÜĴ	1	†	1			1		Г	<u> </u>	1						
Endosulfan II	3.3	45	บัง	55	IJ	i	1	1	1		1	1			Ī		T	1			Π
4,4'-DDD	3.3				Ū·		1	1	1	1	1	1	1	1				1	1		T
Endosulfan sulfate	3.3	30	บ็ป	68	ŬJ	<del> </del>	<del>                                     </del>	†	<del>                                     </del>	1	1	1					T		1		1
4,4'-DDT	3.3			190	<u>היי</u>	<del> </del>	1	<del>                                     </del>	1-	1	T		1	<del> </del>	1		1		1	1	1
Methoxychlor	17.0		111	180		<del>                                     </del>	1	<del> </del>	1	<del>i -</del>	<del> </del>	<u> </u>	<del>                                     </del>	† ·	1	<u> </u>		1	1	T	T
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	170.0		<del>                                      </del>	250		<del> </del>	╁	<del> </del>	<del> </del> -	<del> </del>	+		┼		1	<del> </del>	1	†	t	1	1
Toxaphene Arochlor—1221	33.0		<del>                                      </del>	100		<del> </del>	╁─	<del>  -</del>	╂	<del> </del>	+		┼	<del> </del>	+		+		t	<u> </u>	╁╌
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Arochlor - 1232	67.0			68		1	+	<del> </del>	┨—	+	+-		┼		+	<del> </del>	╁	<del>                                     </del>	┼	<del> </del>	+
Arochlor-1242/1016	33.0			100		+	+	<del> </del>	+	<del>                                     </del>	+		<del> </del>		+-	<del> </del>	+	<del> </del>	<del> </del>	<del> </del>	╁
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Arochlor 1254	33.0			7500	<del> </del>	<del> </del>	+—	<del> </del>	+-	<del> </del>	╁	1	$\vdash$	<del></del>	+	<del> </del>	+	1	+-	<del> </del>	+
Arochlor – 1260	33.0	1800	J	/500		1	Щ.	.1	1	<u> </u>	ᆚ	<u> </u>	ــــــــــــــــــــــــــــــــــــــ	1	_l	1		٠	.1	2JK s'	

# APPENDIX F PESTICIDE/PCB VALIDATED LABORATORY REPORT FORMS

# PESTICIDE ORGANICS ANALYSIS DATA SHEET

EPA SAMPLE NO. BHI-00405

Lab Name: <u>QUANTERRA, MO</u> Contract:	550.56	BODM12 Rev. 00	
Lab Code: ITMO Case No.: SAS No	.: SDG No.:	W0429	
Matrix: (soil/water) SOIL	Lab Sample ID:	7344-002	
Sample wt/vol: 30.0 (g/ml) g	Lab File ID:		
Level: (low/med) LOW	Date Sampled :	01-17-95	
% Moisture: not dec. 44 dec.	Date Extracted: _	01-20-95	
Extraction: (SepF/Cont/Sonc) _SONC_	Date Analyzed:	01-25-95	
GPC Cleanup: (Y/N) N pH:	Dilution Factor:	1	
CAS NO. Compound (ug/I	ENTRATION UNITS: L or ug/Kg) <u>ug/Kg</u>	Q	
319-84-6	3.5 5.3 4.5 1.8 8.2 49 8.3 1.2 25 140 45 6.5 39 170 100 54 8.3 140 59	リープ ファ	ת שלו חחת חות
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U: Concentration of analyte is less than the value given.

SD: See dilution

Elevated detection limit due to PCB interference.
FORM I PEST

1/87 Rev.

PESTICIDE ORGANICS ANALYSIS DATA SPECTIO 1749

EPA SAMPLE NO.

RHI-00405

Lab Name: OUANTERRA, MO Contract:	550.56	BODMT3 Rev. 00
Lab Code: <u>ITMO</u> Case No.:	SAS No.: SDG No.:	W0429
Matrix: (soil/water)SOIL	Lab Sample ID:	7344-004
Sample wt/vol: <u>30.0</u> (g/ml) <u>q</u>	Lab File ID:	
Level: (low/med) LOW	Date Sampled :	01-17-95
% Moisture: not dec. 68 dec.	Date Extracted: _	01-20-95
Extraction: (SepF/Cont/Sonc) SONC	Date Analyzed:	01-25-95
GPC Cleanup: (Y/N) N pH:	Dilution Factor:	1
CAS NO. Compound	CONCENTRATION UNITS: (ug/L or ug/Kg) ug/Kg	Q
319-84-6	6 . 2 9 . 3 4 . 1 3 . 1 36 86 14 2 . 1 82 440 55 24 8e 68 190 180 110	
53469-21-9/12674-11-2-Aroclor-1242/1016 12672-29-6	100	

U: Concentration of analyte is less than the value given.

SD: See dilution

X: Elevated detection limit due to PCB interference.
FORM I PEST

F8 4/30/95

1/87 Rev.

BHI-00405 Rev. 00

#### 1D PESTICIDE ORGANICS ANALYSIS DATA SHEET

EPA SAMPLE NO.

Lab Name: QUANTERRA,MO Contract: 550-56	BODMT4
Lab Code: ITMO Case No.: SAS No.: SDG No.	: _W0430
Matrix: (soil/water) <u>SOIL</u> Lab Sample ID:	7444-008
Sample wt/vol: 30.0 (g/ml) g Lab File ID:	
Level: (low/med) LOW Date Sampled:	01-27-95
% Moisture: not dec. 6 dec. Date Extracted:	02-03-95
Extraction: (SepF/Cont/Sonc) SONC Date Analyzed:	02-08-95
GPC Cleanup: (Y/N) N pH: Dilution Factor:	1
CAS NO. Compound (ug/L or ug/Kg) UG/KG	Q
319-84-6alpha-BHC	
11097-69-1Aroclor-1254 36 11096-82-5Aroclor-1260 36	

U: Concentration of analyte is less than the value given.



FORM I PEST

# APPENDIX G METALS DATA SUMMARY TABLES

Project: WESTING	IOUSE-	HANFOR	1D	]																	
Laboratory: Quante				]																	
Case	SDG: W	0429		]																	
Sample Number		BODMT	2	BODMT3	l									Ĭ		<u> </u>					
Location		Test Pit	#2	Test Pit	#2	]												1			$\neg \neg$
Remarks	MG/KG	Soil		Soil												1					
Sample Date		01/17/	95	01/17/9	5												-	İ .			$\neg \neg$
Inorganic Analytes		Result	a	Result	Q	Result	O	Result	a	Result	Q	Result	Q	Result	Q	Result	Q	Result	Q	Result	Q
Aluminum	20	53700	l	63400			L		<u> </u>		<u> </u>								<u>l</u>	]	
Antimony	10	7.8		10.1	U		L		<u> </u>								<u> </u>				
Arsenic	1	14.4	ļ	41.1				<u> </u>	<u> </u>	<u> </u>	<u> </u>				1				L		
Barlum	20	74.3		107		ļ. <u></u>	<u> </u>		<u> </u>										Г		
Beryllium	0.7	0.59	-	0.76						<u> </u>	]						ļ				
Cadmium	2	2.6	_	4.7	·									i							П
Calcium	500	3630		4200					L		<u> </u>			ļ							
Chromium	2	66.4	L	77.1															Ì		$\Box$
Cobalt	5	7.3		9.4		L	Ι.		]							1			†		П
Copper	2.5	68.6		55.0					Γ									1			П
iron	10	20100		21000							1			<u> </u>	1	<u> </u>	<del>                                     </del>				1
Lead	0.3	104		197							1				1	ļ	1-				
Magnesium	500	3090	İ	4180	1		$\Box$				<u> </u>	1			1		<del>                                     </del>	<b> </b>	t —		
Manganese	1.5	140		201								<del> </del>		,	1		$\Box$		1		
Nickel	4	14.7	$\Box$	18.8							1	1			<u> </u>		$\vdash$		ļ		
Potassium	500	654		970							İ	<u> </u>			<del>                                     </del>		<u> </u>	<u> </u>	t	<del></del>	╁
Selenium	0.5	0.18	U	0.31	Ū				<u> </u>						1	<del></del>	<del>                                     </del>		<del>                                     </del>		$\vdash$
Silver	20	0.76		1.3				† · · · · · · · · · · · · · · · · · · ·		<del>                                     </del>	<b>†</b>	· · · · ·			1		1		╁──		
Sodium	100	244		286	П		1-								1		<del>                                     </del>		一	<del></del>	Н
Thallium	1	0.28	T	0.32			П		Г		İ				<b>†</b>	<u> </u>	<u> </u>		<del>                                     </del>		H
Vanadium	5	45.0		46.5					Г		<b> </b>			<del></del> -	1		<del>                                     </del>	† <del></del>	<del>                                     </del>	· · · · · · · · · · · · · · · · · · ·	H
Zinc	2	450		1030				1	1						一	†···	T	<del>                                     </del>	✝		
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# APPENDIX H METALS VALIDATED LABORATORY REPORT FORMS

Rev. 00 EPA SAMPLE NO.

# INORGANIC ANALYSES DATA SHEET

Concentration Units (ug/L or mg/kg dry weight): MG/KG

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CAS No.	Analyte	Concentration	c	Q	м
7429-90-5	Aluminum	53700	-	} <del></del>	P
7440-36-0	Antimony	7.8	B		$\bar{P}^-$
7440-38-2	Arsenic	14.4	В		$ \mathbf{F}^{-} $
7440-39-3	Barium	74.3			$\mathbf{P}^{-}$
7440-41-7	Beryllium	0.59	B	<del></del>	$ \bar{P}^- $
7440-43-9	Cadmium	2.6	В		$ \bar{\mathbf{p}}^{-} $
7440-70-2	Calcium	3630	1		P_
7440-47-3	Chromium	66.4			P-
7440-48-4	Cobalt	7.3	B		$\mathbf{p}^{-}$
7440-50-8	Copper	68.6			P_
7439-89-6	Iron	20100	—		P
7439-92-1	Lead	104			P_
7439-95-4	Magnesium	3090	_		$P^{-1}$
7439-96-5	Manganese	140			P_
7440-02-0	Nickel	14.7			P_
7440-09-7	Potassium	654	$\overline{\mathbf{B}}$		P_
7782-49-2	Selenium_	0.18	U		F_
7440-22-4	Silver	0.76	В		P_
7440-23-5	Sodium	244			P_
7440-28-0	Thallium	0.28	B		F_
7440-62-2	Vanadium_	45.0	_		P_
7440-66-6	Zinc	450	_		P_
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nments:		
	FORM I - IN	275 SW-846

U.S. EPA - CLP

BHI-00405 Rev. 00 EPA SAMPLE NO.

# INORGANIC ANALYSES DATA SHEET

BODMT4 ab Name: QUANTERRA MO ab Code: ITMO Case Contract: 550.56 Case No.: SAS No.: SDG No.: W0430 atrix (soil/water): SOIL\_ Lab Sample ID: 7444-008 evel (low/med): LOW Date Received: 01/30/95 93.5 Solids: Concentration Units (ug/L or mg/kg dry weight): MG/KG CAS No. Analyte Concentration | C M Q 7429-90-5 Aluminum 4010 P Antimony 6.3 E ₽\_ BJ 7440-36-0 F 7440-38-2 Arsenic 0.68 B P Barium 93.5 7440-39-3 7440-41-7 Beryllium 0.45 B  $\mathbf{P}^{-}$ RZ Cadmium 7440-43-9 0.41 U  $\mathbf{p}^{-}$ Calcium -4020  $\mathbf{P}^{\mathsf{T}}$ 7440-70-2 P 7440-47-3 Chromium 4.9 7440-48-4 Cobalt 13.0 P 7440-50-8 Copper 14.5  $\mathbf{p}^{\mathsf{T}}$ 7439-89-6 Iron 13400 Þ. 7439-92-1 Lead 1.7 F 7439-95-4 Magnesium 4060  $\mathbf{P}^{-}$ 7439-96-5 Manganese 311  $\mathbf{P}^{-}$ J 7.1 7440-02-0 Nickel P 7440-09-7 Potassium 477 B P  $\overline{0}.11|U$ F 7782-49-2 Selenium 7440-22-4 Silver 0.41 U  $\mathbf{P}^{\mathsf{T}}$ 7440-23-5 Sodium 129  $\mathbf{P}$ Thallium 7440-28-0 Ō.10₺ F 107 Vanadium -7440-62-2 68.7  $\mathbf{p}^{-}$ 7440-66-6 Zinc 56.2 Clarity Before: Color Before: Texture: Clarity After: Color After: Artifacts: comments:

SW-846

FORM I - IN

# APPENDIX I GENERAL CHEMISTRY DATA SUMMARY TABLES

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Project: WESTINGHOU	JSE-HAN	FORD																			
Laboratory: QUANTER				]																	
Case	SDG: W	0429		1																	
Sample Number		BODMT2	2	BODMT3	)	1		1		1		Τ		T				T		<del></del>	—
Location		Test Pit	#2	Test Pit	#2	1				1		†		<del> </del> -		<del> </del>		<del> </del>		ļ ———	
Remarks	UG/G	Soll		Soil		<u> </u>				1		·		<del>                                     </del>		<del> </del> -		<del> </del>		<del> </del>	
Sample Date		01/17/9	5	01/17/9	5	†—— <u> </u>		1		†		<del> </del>		<del> </del>				<del>                                     </del>		ļ <u>.</u>	
Wet Chemistry Analyte	s   Method	Result	Q	Result	Q	Result	Q	Result	Q	Result	Q	Result	O	Result	TO.	Regist	a	Regist	10	D-014	75
7 1401140	300.0	1.70	יוי	0.23	1	I				1		· · · · · · · · · · · · · · · · · · ·	1	1	+=		<u> </u>	IIIOOGIL	-	1100 CHI	+
Sulfate	300.0	2230		4040			l				<b>†</b>	<u> </u>		<del> </del>	一	<del> </del>	!-	<del>                                     </del>	┼┈		╁
Chloride	300.0			20.3				1	T	<u> </u>	!			<del> </del>	<del> </del>		1	<del> </del>	<del> </del>		+
Sulfide	9030	21.0		44.7			1				1 -	<u> </u>		<del> </del>	┰	<del>                                     </del>	$\vdash$	<del> </del>	┰		╌
Nitrate	300.0			50.3			_		$T^-$	<u> </u>	t	<b>1</b>		t	†		1	<del> </del>	<del> </del>		+
Nkrite	300.0			0.61	U				<b>—</b>	·	1		_		<del>                                     </del>	<del></del>	╁	<del> </del>	<del>├</del>		+-
Phosphate	300.0			30.6	Ü				1	<del>                                     </del>	$\vdash$	† <u>-</u>		<del>                                     </del>	$\dagger$	<del> </del>	<del> </del>	<del> </del>	╂─		+-
Nitrate/Nitrite	353.1	53.4		35.4				1	1		<del>                                     </del>	<u> </u>	-	<del>                                     </del>	$\vdash$	<del></del> -	<del> </del>	<del> </del>	<del>                                     </del>		+-
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RUC for RJS 8 BHI 000

## APPENDIX J

### GENERAL CHEMISTRY VALIDATED LABORATORY REPORT FORMS

P.O. Box 1970 Richland, WA 99352

Project: 550.56

Category: Mitrate Method: EPA 300.0 Matrix: SOLID

Emironmental Services BHI-00405 Rev. 00

Sample Date : 01/17/95 Receipt Date : 01/18/95 Report Date : 03/01/95

Client ID	<b>Quanterra</b> ID	Analyte	CAS Number	Blank Sample Name	Prep. Date	Analyses Date	Result	Unit	Qual.	Detection Limit	n Dil
BODMT6	7344-001	Nitrate	14797-55-8	QCBLK57444-2	01/20/9	5 01/20/95	0.20	UG/G	U	0.20	1
BODMT6	7344-001DUP	Nitrate	14797-55-8	QCBLK57444-2	01/20/9	5 01/20/95	0.20	UG/G	U	0.20	1
BODMT6	7344-001MS	Nitrate	14797-55-8	OCBLK57444-2	01/20/9	5 01/20/95	106	<b>X</b> REC			1
BODMT2	7344-002	Nitrate	14797-55-8	QCBLK57444-2	01/20/9	5 01/20/95	69.5	UG/G		1.70	5
BODMT3	7344-004	Nitrate	14797-55-8	OC8LK57444-2	01/20/9	5 01/20/95	50.3	UG/G		1.22	2
NA	QCBLK57444-2	Nitrate	14797-55-8	QCBLK57444-2	01/20/9	5 01/20/95	0.20	UG/G	ช	0.20	1
NA	QCLCS57444-2	Nitrate	14797-55-8	QCBLK57444-2	01/20/99	5 01/20/95	100	XREC			1

R15 ps (4/1/85

Quanterra-Richland 1 760 P.O. Box 1970 Richland, UA 99352

Project: 550.56

. Environmental

Service BHI-00405 Rev. 00

Category: Orthophosphate Method: EPA 300.0 Matrix: SOLID

Sample Date : 01/17/95 Receipt Date : 01/18/95 Report Date : 03/01/95

Client ID	Quanterra ID	Analyte	CAS Number	Blank Sample Name	Prep. Date	Analyses Date	Result	Unit	Qual.	Detection Limit	Dil
BODMT6	7344-001	Ortho-Phosphate	7778-77-0	QCBLX57444-2	01/20/9	01/20/95	9-86	UG/G	Ü	9.86	1
BODMT6	7344-001DUP	Ortho-Phosphate	7778-77-0	QCBLK57444-2	01/20/9	01/20/95	9_84	UG/G	U	9.84	1
BODMT6	7344-001MS	Ortho-Phosphate	7778-77-0	QCBLK57444-2	01/20/9	01/20/95	102	<b>X</b> REC			1
BODMT2	7344-002	Ortho-Phosphate	7778-77-0	QCBLK57444-2	01/20/95	01/20/95	17.0	UG/G	U	17.0	1
BODMT3	7344-004	Ortho-Phosphate	7778-77-0	QCBLK57444-2	01/20/95	01/20/95	30.6	UG/G	U	30.6	1
NA ·	QCBLK57444-2	Ortho-Phosphate	7778-77 <b>-</b> 0	QCBLK57444-2	01/20/95	01/20/95	10.0	UG/G	บ	10.0	1
NA	QCLCS57444-2	Ortho-Phosphate	7778-77-0	QCBLK57444-2	01/20/95	01/20/95	96	<b>X</b> REC			1

PBC 15

P.O. Box 1970 Richland, WA 99352

Project: 550.56

Category: Fluoride Method: EPA 300.0 Matrix: SOLID

Contract Contract Service BHI-00405 Rev. 00

Sample Date : 01/17/95 Receipt Date : 01/18/95 Report Date : 03/01/95

Client ID	Quanterra ID	Analyte	CAS Number	Blank Sample Name	Prep. Date	Analyses Date	Result	Unit	Qual.	Detection Limit	n Dil
BODMT6	7344-001	Fluoride	16984-48-8	QCBLK57444-2	01/20/9	5 01/20/95	0.99	UG/G	Ų	0.99	1
BODMT6	7344-001DUP	Fluoride	16984-48-8	QCBLK57444-2	01/20/9	5 01/20/95	0.98	UG/G	U	0.98	1
BODMTS	7344-001MS	Fluoride	16984-48-8	QCBLK57444-2	01/20/9	5 01/20/95	107	<b>X</b> REC			1
BODMT2	7344-002	Fluoride	16984-48-8	QCBLK57444-2	01/20/9	5 01/20/95	1.70	UG/G	U	1.70	1
BODMT3	7344-004	Fluoride	16984-48-8	QCBLK57444-2	01/20/95	5 01/20/95	6.25	UG/G		3.06	1
NA	QCBLK57444-2	Fluoride	16984-48-8	QCBLK57444-2	01/20/95	01/20/95	1.00	UG/G	U	1.00	1
NA	QCLC\$57444-2	Fluoride	16984-48-8	QCBLK57444-2	01/20/95	01/20/95	103	<b>X</b> REC			1

RDC 1-715

MI 1960. 1762

Quanterra-Richland P.O. Box 1970 Richland, WA 99352

Project: 550.56

Category: TOX
Method: EPA 9020
Matrix: SOLID

**@**uanterra

Environmental BHI-00405

Rev. 00

Sample Date : 01/17/95 Receipt Date : 01/18/95 Report Date : 03/01/95

<u> </u>								
Client ID	Quanterra ID	Analyte	CAS Number	Blank Sampie Name	Prep. Analyses Date Date	Result Unit	Qual.	Detection Limit Di
BODMT6	7344-001	EOX	IY-EOX	QCBLK58657-1	02/06/95 02/06/95	47.1 UG/G	U	47_1
BODMT6	7344-001DUP	EOX	IT-EOX	QCBLK58657-1	02/06/95 02/06/95	46.1 UG/G	U	46.1
BODMT6	7344-001MS	EOX	IT-EOX	QCBLK58657-1	02/06/95 02/06/95	101 XREC		
BODMT2	7344-002	EOX	IT-EOX	QCBLK58657-1	02/06/95 02/06/95	86.8 UG/G	U	86.8
BODMT3	7344-004	EOX	IT-EOX	QCBLK58657-1	02/06/95 02/06/95	150 UG/G	U	150
NA	QCBLK58657-1	EOX	IT-EOX	QCBLK58657-1	02/06/95 02/06/95	50.0 UG/G	U	50.0
NA	QCLCS58657-1	EOX	IT-EOX	QCBLK58657-1	02/06/95 02/06/95	99 XREC		

Quanterra-Richland 30 1 2703 P.O. Box 1970 1 20 1 2703 Richland, WA 99352

Project: 550.56

Category: NO2-NO3 Method: EPA 353.1 Matrix: SOLID

Sample Date : 01/17/95 Receipt Date : 01/18/95 Report Date : 03/01/95

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Rev. 00

Client ID	Quanterra ID	Analyte	CAS Number	Blank Sample Name	Prep. Date	Analyses Date	Result	Unit	Qual.	Detection Limit	n Dil.
					2010	-	NGSQ. C	0.,,,	-uar.	C FIGURE	DIL.
BODMT6	7344-001	Nitrate/Nitrite	C-005	QCBLK57497-2	01/24/9	5 01/24/95	0.48	UG/G	U	0.48	1
BODMT6	7344-001DUP	Witrate/Witrite	C-005	QCBLK57497-2	01/24/9	01/24/95	0.50	UG/G	U	0.50	1
BODMT6	7344-001MS	Witrate/Witrite	C-005	QC8LK57497-2	01/24/9	5 01/24/95	104	XREC			1
BODMT2	7344-002	Witrate/Witrite	C-005	QC8LK57497-2	01/24/9	5 01/24/95	53.4	UG/G		4.40	5
BODMT3	7344-004	Nitrate/Nitrite	C-005	QCBLK57497-2	01/24/9	01/24/95	35.4	UG/G		3.09	2
NA	QCBLK57497-2	Nitrate/Nitrite	C-005	QCBLK57497-2	01/24/9	01/24/95	0.50	UG/G	U	0.50	1
4A	QCLCS57497-2	Nitrate/Nitrite	C-005	QCBLK57497-2	01/24/95	01/24/95	92	%REC			1

RBC for RJS 4/7/95

Quanterra-Richland 39 764 P.O. Box 1970 Richland, WA 99352

Project: 550.56

BHI-00405 Rev. 00

Category: Nitrite Method: EPA 300.0 Matrix: SOLID

Sample Date : 01/27/95 Receipt Date : 01/30/95 Report Date : 03/07/95

Client	Quanterra			Blank Sample	Prep.	Analyses				Detection	
ID	ID	Analyte	CAS Number	Name	Date	Date	Result	Unit	Qual.	Limit	Dil.
80DMS4	7444-001	Nitrite	7632-00-0	QCBLK58660-1	02/03/99	02/03/95	0.21	UG/G	U	0.21	1
BODMS4	7444-001DUP	Nitrite	7632-00-0	QCBLK58660-1	02/03/99	02/03/95	0.21	UG/G	U	0.21	1
BODMS4	7444-001MS	Nitrite	7632-00-0	QCBLK58660-1	02/03/95	02/03/95	103	<b>X</b> REC			5
BODMT4	7444-008	Nitrite	7632-00-0	QCBLK58660-1	02/03/95	02/03/95	0.19	UG/G	U	0.19	1
NA	QCBLK58660-1	Nitrite	7632-00-0	QCBLK58660-1	02/03/95	02/03/95	0.20	UG/G	U	0.20	1
NA	QCLCS58660-1	Nitrite	7632-00-0	QCBLK58660-1	02/03/95	02/03/95	98	<b>X</b> REC			1

phc for JS

Quanterra-Richtand 765 P.O. Box 1970 Richtand, WA 99352

Project: 550.56

Category: Chloride Method: EPA 300.0 Matrix: SOLID

BHI-00405 Rev. 00

Sample Date : 01/27/95 Receipt Date : 01/30/95 Report Date : 03/07/95

Client ID	Quanterra 1D	Analyte	CAS Number	Blank Sample Name	Prep. Analyses Date Date	Result Uni	Qual.	Detectio Limit	n Dil.
80DMS4	7444-001	Chloride	16887-00-6	QCBLK58660-1	02/03/95 02/03/95	2.59 UG/0	υ	2.59	1
BODMS4	7444-001DUP	Chloride	16887-00-6	QCBLK58660-1	02/03/95 02/03/95	2.58 UG/0	υ	2.58	1
B001/54	7444-001MS	Chloride	16887-00-6	QCBLK58660-1	02/03/95 02/03/95	_100 %REG	;		5
BODMT4	7444-008	Chloride	16887-00-6	QCBLK58660-1	02/03/95 02/03/95	2.40 UG/0	. U	2.40	1
NA	QCBLK58660-1	Chloride	16887-00-6	QCBLK58660-1	02/03/95 02/03/95	2.50 UG/0	i U	2.50	1
NA	QCLCS58660-1	Chloride	16887-00-6	QCBLK58660-1	02/03/95 02/03/95	90 %REC			1

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Quanterra-Richtand P.O. Box 1970 Richland, WA 99352

Project: 550.56

Category: Sulfate Method: EPA 300.0 Matrix: SOLID

BHI-00405 Rev. 00

Sample Date : 01/27/95 Receipt Date : 01/30/95 Report Date : 03/07/95

Client	Quanterra			Blank Sample		Analyses				Detection	
ID	10	Analyte	CAS Number	Kame	Date	Date	Result	Unit	Qual.	Limit	Dil.
BODMS4	7444-001	Sulfate	14808-79-8	QCBLK58660-1	02/03/9	02/03/95	25.1	UG/G		10.4	1
BODMS4	7444-001DUP	Sulfate	14808-79-8	QCBLK58660-1	02/03/95	02/03/95	24.7	UG/G		10.3	1
BODMS4	7444-001MS	Sulfate	14808-79-8	QCBLK58660-1	02/03/99	02/03/95	95	<b>X</b> REC			5
BODMT4	7444-008	Sulfate	14808-79-8	QCBLK58660-1	02/03/95	02/03/95	54.1	UG/G		9.60	1
NA	QCBLK58660-1	Sulfate	14808-79-8	QCBLK58660-1	02/03/99	02/03/95	10.0	UG/G	U	10.0	1
NA	QCLCS58660-1	Sulfate	14808-79-8	QCBLK58660-1	02/03/99	02/03/95	92	<b>XREC</b>			1

9-1-356.1767

Quanterra-Richland P.O. Box 1970 Richland, WA 99352

Project: 550.56

Category: NO2-NO3
Method: EPA 353.1
Matrix: SOLID

Sample Date : 01/27/95 Receipt Date : 01/30/95 Report Date : 03/07/95

BHI-00405

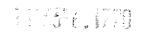
Rev. 00

Client 1D	Quanterra ID	Analyte	CAS Number	Blank Sample Name	Prep. Date	Analyses Date	Result	Unit	Qual.	Detection Limit	Dil.
BODMS4	7444-001	Nitrate/Nitrite	C-005	QCBLK59558-1	02/15/9	5 02/15/95	4.73	UG/G		0.51	1
BODMS4	7444-001DUP	Nitrate/Nitrite	C-005	QCBLK59558-1	02/15/95	5 02/15/95	4.75	UG/G		0.51	1
BODMS4	7444-001MS	Nitrate/Nitrite	C-005	QC8LK59558-1	02/15/9	02/15/95	80	%REC			1
BODMT4	7444-008	Nitrate/Nitrite	C-005	QCBLK59558-1	02/15/9	5 02/15/95	1.00	UG/G		0.53	1
NA	QCBLK59558-1	Nitrate/Nitrite	C-005	QCBLK59558-1	02/15/95	02/15/95	0.50	UG/G	U	0.50	1
NA	QCLCS59558-1	Nitrate/Nitrite	C-005	QCBLK59558-1	02/15/95	02/15/95	107	ZREC			1

# APPENDIX K RADIOCHEMISTRY DATA SUMMARY TABLES

Laboratory: QES			1																
	SDG:																		
Sample Number		B0DMT2	BODMT3			ļ								L					
Location		Test Pit #2	Test Pit #2							l									
	pCi/g	Soil	Soil																
Sample Date		01/17/95	01/17/95	Í															
Radiochemistry Analysis		Result Q	Q	Result	Q	Result	Q.	Result	Q	Result	a	Result	Q	Result	Q	Result	Q	Result	ŢQ
Gross Alpha		9.95	25.0		<u>.  </u>	.l	ļ			L	<u></u>					<u></u>	<u>L</u>		
Gross Beta		20.6	21.1	<u> </u>	.i	<u> </u>	ļ		L	<u> </u>									L
Cobalt – 58		0.0165 U	0.0143 U										l		_				T
Cobalt – 60		0.929	2.43	<u> </u>	<u> </u>							l					$I^-$		$\top$
Cesium – 137DA		2.72	3.09	<u> </u>			L			L					[				1
Europium – 152		1.55	1.96																T
Europium – 154		0.331	0.46	Ī	ĺ				i					i	1	<u> </u>			1
Europlum – 155		0.0790 U	0.112 U		"					i					† <u> </u>				1
lron – 59		- 0.0454 U	- 0.0669 U	1	1										1	<del> </del>	!		+-
Potassium – 40		9.57	6.77		$\vdash$				T					i — — —	┿	<b></b>			+
Radium - 224DA		0.863	1.05											<del> </del>	<del> </del>	<del> </del>	$\vdash$		+-
Radium - 226DA		0.813	0.801	1	1		_					<b></b>		<del></del>	<del> </del>	<del>                                     </del>	<del> </del>		+
Radium - 228DA	•	0.721 J	1.51 J	†——	-		<del>                                     </del>				L	<b> </b>	<del>                                     </del>	<del> </del>	<del> </del>	<del>                                     </del>	<del> </del>	· · · · · · · · ·	+-
Uranium-238DLP	·	2.76 J	5.90 J		-		<u> </u>								┼	<del> </del>			+
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BHI-00405 Rev. 00



# APPENDIX L

### RADIOCHEMISTRY VALIDATED LABORATORY REPORT FORMS

### **SAMPLE RESULTS**

LAB NAME:

ITAS-RICHLAND

SDG:

W0429

LAB SAMPLE ID:

50132303

**MATRIX:** 

SOIL

**CLIENT ID:** 

B0DMT2

DATE RECEIVED:

1/18/95

ISOTOPE		OUNTING RROR (2s)	TOTAL ERROR (2s)	MDA	REPORT UNIT	YIELD	METHOD NUMBER
CO-58	1.65E-02 U	2.6E-02	2.6E-02	4.42E-02	pCi/g	N/A	RD3219
CO-60	9.29E-01	6.0E-02	1.1E-01	N/A	pCi/g	N/A	RD3219
CS-137DA	2.72E+00	7.0E-02	2.8E-01	N/A	pCi/g	N/A	RD3219
EU-152	1.55E+00	1.2E-01	1.9E-01	N/A	pCi/g	N/A	RD3219
EU-154	3.31E-01	6.6E-02	7.4E-02	1.36E-01	pCi/g	N/A	RD3219
EU-155	7.90E-02 U	5.9E-02	6.0E-02	8.92E-02	pC <b>i/</b> g	N/A	RD3219
FE-59	-4.54E-02 U	6.8E-02	6.8E-02	1.07E-01	pCi/g	N/A	RD3219
K-40	9.57E+00	5.7E-01	1.1E+00	N/A	pCi/g	N/A	RD3219
RA-224DA	8.63E-01	4.5E-02	9.7E-02	N/A	pCi/g	N/A	RD3219
RA-226DA	8.13E-01	7.0E-02	1.1E-01	N/A	pCi/g	N/A	RD3219
RA-228DA	7.21E-01 丁	1.3E-01	1.5E-01	N/A	pCi/g	N/A	RD3219
U-238DLP	2.76E+00 J	1.2E+00	1.2E+00	N/A	pCi/g	N/A	RD3219
ALPHA	9.95E+00	4.8E+00	4.9E+00	5.93E+00	pCi/g	100.00%	RD3222
BETA	2.06E+01	3.3E+00	3.6E+00	3.58E+00	pCi/g	100.00%	RD3222

Number of Results: 14

X18 95

BHI-00405 Rev. 00

### **SAMPLE RESULTS**

LAB NAME:

ITAS-RICHLAND

SDG:

W0430

LAB SAMPLE ID:

50151812

**MATRIX:** 

SOIL

**CLIENT ID:** 

B0DMT4

DATE RECEIVED:

1/30/95

ISOTOPE		OUNTING RROR (2s)	TOTAL ERROR (2s)	MDA	REPORT UNIT	YIELD	METHOD NUMBER
CO-58	4.35E-03 ∪	1.2E-02	1.2E-02	2.06E-02	pCi/g	N/A	RD3219
CO-60	1.19E-02 ∪	1. <b>0</b> E-02	1.0E-02	1.90E-02	pCi/g	N/A	RD3219
CS-137DA	2.54E-02	1.4E-02	1.4E-02	N/A	pCi/g	N/A	RD3219
EU-152	1.86E-02 <i>U</i>	2.6E-02	2.6E-02	4.23E-02	pCi/g	N/A	RD3219
EU-154	-5.84E-03 ∪	3.1E-02	3.1E-02	5.33E-02	pCi/g	N/A	RD3219
EU-155	3.01E-02 ∪	2.2E-02	2.3E-02	3.84E-02	pCi/g	N/A	RD3219
FE-59	-4.77E-02 ∪	3.3E-02	3.4E-02	5.04E-02	pCi/g	N/A	RD3219
K-40	9.42E+00	4.3E-01	1.0E+00	N/A	pCi/g	N/A	RD3219
RA-224DA	4.14E-01	2.8E-02	5.0E-02	N/A	pCi/g	N/A	RD3219
RA-226DA	3.67E-01	<b>4.0E-</b> 02	5.5E-02	N/A	pCi/g	N/A	RD3219
RA-228DA	4.42E-01	6. <b>7</b> E-02	8.0E-02	N/A	pCi/g	N/A	RD3219
ALPHA	1.90E+00 U	2.7E+00	2.7E+00	5.34E+00	pCi/g	100.00%	RD3222
BETA	1.59E+01	3.0E+00	3.2E+00	3.66E+00	pCi/g	100.00%	RD3222

Number of Results: 13

RBC 38-3-95

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